

Product Sheet

H_HER2 EMT6 Cell Line

Catalog number: GM-C33821

Version 3.3.1.260819

Description	H_HER2 EMT6 Cell Line is a clonal stable EMT6 cell line that constitutively expresses the human HER2 gene, constructed using lentiviral technology.
Quantity	5E6 Cells per vial, 1 mL
Product Format	1 vial of frozen cells
Shipping	Shipped on dry ice
Storage Conditions	Liquid nitrogen immediately upon receipt
Target	Human_HER2
Gene ID/Uniprot ID	P04626-1
Host Cell	EMT6
Recovery Medium	Waymouth's MB 752/1+15% FBS+1% P.S
Growth medium	Waymouth's MB 752/1+15% FBS+1% P.S+1.5 µg/mL Puromycin
Note	None
Freezing Medium	90% FBS+10% DMSO
Growth properties	Adherent
Growth Conditions	37°C, 5% CO ₂
Mycoplasma Testing	The cell line has been screened to confirm the absence of Mycoplasma species.
Safety considerations	Biosafety Level 2
Note	It is recommended to expand the cell culture and store a minimum of 10 vials at an early passage for potential future use.

Materials

Reagent	Manufacturer/Catalogue No.
Waymouth's MB 752/1	Procell/PM151215
Fetal Bovine Serum	ExCell/FSP500
Pen/Strep	Thermo/15140-122
Puromycin	Genomeditech/ GM-040401
Anti-H ₂ HER2 hIgG1 Antibody(Margetuximab)	Genomeditech/ GM-49468AB

Figures

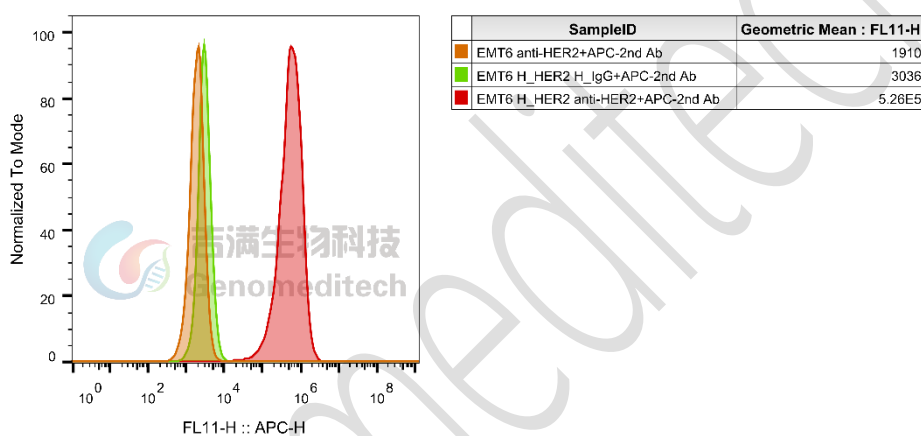


Figure 1 | H₂HER2 EMT6 Cell Line (Cat. GM-C33821) was determined by flow cytometry using Anti-H₂HER2 hIgG1 Antibody(Margetuximab)(Cat. [GM-49468AB](#)).

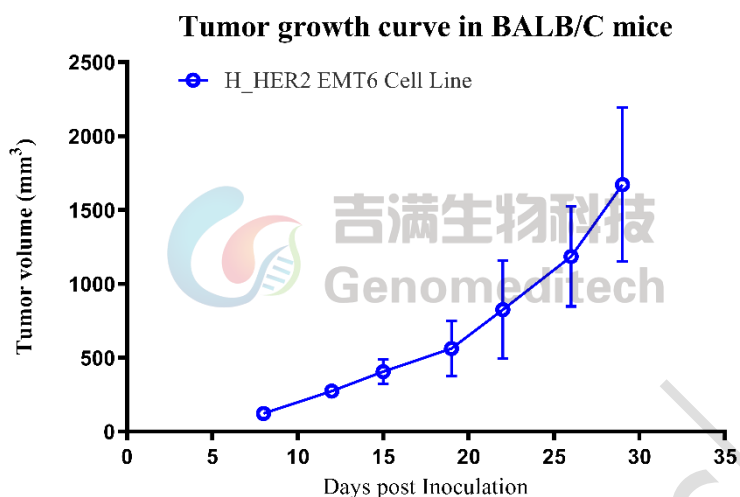


Figure 2 | Tumor growth curves of H_HER2 EMT6 in BALB/C mice. H_HER2 EMT6 cells (1×10^6 per mouse) were subcutaneously inoculated into BALB/C mice (female, 8 weeks old, $n = 3$). Tumor volume was measured twice per week and is presented as mean $\pm$ SEM.

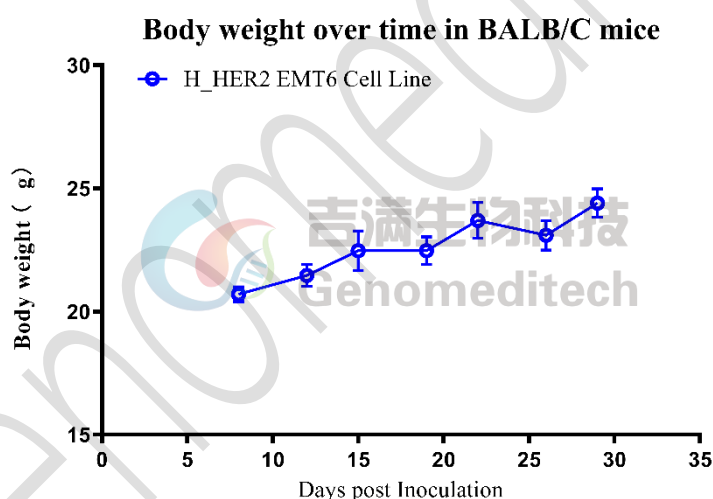


Figure 3 | Body weight changes after implantation of H_HER2 EMT6 in BALB/C mice. Under the same conditions, body weight was measured twice per week and is presented as mean $\pm$ SEM.

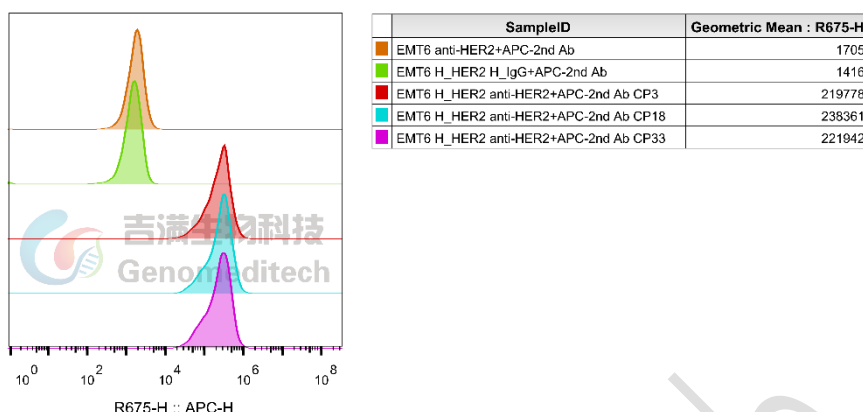


Figure 4 | The passage stability of the H_HER2 EMT6 Cell Line (Cat. GM-C33821) was determined by flow cytometry using Anti-H_HER2 hIgG1 Antibody(Margetuximab)(Cat. [GM-49468AB](#)).

Cell Recovery

Recovery Medium: Waymouth's MB 752/1+15% FBS+1% P.S

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

- Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 - 3 minutes).
- Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
- Transfer the vial contents to a centrifuge tube containing 5.0 mL complete culture medium and spin at approximately 176 x g for 5 minutes. Discard supernatant.
- Resuspend cell pellet with the recommended recovery medium. And dispense into appropriate culture dishes.
- Incubate the culture at 37°C in a suitable incubator. A 5% CO₂ in air atmosphere is recommended if using the medium described on this product sheet.

Cell Freezing

Freezing Medium: 90% FBS+10% DMSO

- Centrifuge at 176 x g for 3 minutes to collect cells.
- Resuspend the cells in pre-cooled freezing medium and adjust the cell density to 5E6 cells/mL.
- Aliquot 1 mL into each vial.
- Place the vial in a controlled-rate freezing container and store at -80°C for at least 1 day, then transfer to liquid nitrogen as soon as possible.

Cell passage

Growth medium: Waymouth's MB 752/1+15% FBS+1% P.S+1.5 µg/mL Puromycin

For the first 1 to 2 passages post-resuscitation, use the recovery medium. Once the cells have stabilized, switch to a growth medium.

- Remove and discard culture medium.
- Briefly rinse the cell layer with PBS to remove all traces of serum that contains trypsin inhibitor.
- Add 1.0 mL of 0.25% (w/v) Trypsin-EDTA solution to dish and observe cells under an inverted microscope until cell layer is dispersed (usually within 30 to 60 seconds at 37°C).
- Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.
- Add 2.0 mL of growth medium to mix well and aspirate cells by gently pipetting.
- After centrifugation, resuspend the pellet and add appropriate aliquots of the cell suspension to new culture vessels.
- Incubate cultures at 37°C.

Subcultivation Ratio: A subcultivation ratio of 1:4 - 1:5 is recommended

Medium Renewal: Every 2 to 3 days

Notes

- None

Sequence

ERBB2(HER2) [P04626-1](#)

MELAAALCRWGLLLALLPPGAASTQVCTGTDMKLRLPASPETHLDMRLHLYQGCQVVQGNLELTYLPTNASL
SFLQDIQEVQGYVLIHNVQRQVPLQRLRIVRGQTQFEDNYALAVLDNGDPLNNTTPVTGASPGGLRELQLR
SLTEILKGGVLIQRNPQLCYQDTILWKDIFHKNNQLALTLIDTNRSRACHPCSPMCKGSRGWGESSEDCQSLT
RTVCAGGCARCKGPLPTDCCHEQCAAGCTGPKHSDCLACLFHNSGICELHCPALVTYNTDTFESMPNPEGR
YTFGASCVTACPYNYLSTDVGSCTLCPLHNQEVTAEDGTQRCEKCSKPCARVCYGLGMEHLREVRAV TSA
NIQEFAGCKKIFGSLAFLPESFDGDPASNTAPLQPEQLQVFETLEEITGYLYISAWPDSLPLDSVFNQLQVIRGRI
LHNGAYSLTLQGLGISWLGLRSLRELGSGLALIHNTHLCFVHTVPWDQLFRNPHQALLHTANRPEDECVGE
GLACHQLCARGHCWGPPTQCVNCSQFLRGQECVEECRVLQGLPREYVNARHCLPCHPECQPQNGSVTCFG
PEADQCVACAHYKDPFPCVARCPGSKPDLSYMPIWKFPDEEGACQPCPINCTHSCVDLDDKGCPAEQRAS
LTSIISAVVGILLVVVLGVVFGILIKRRQKIRKTYMRLLQETELVEPLTPSGAMPNQAQMRILKETELRKVK
VLGSGAFGTVYKGIWIPDGENVKIPVAIKVLRENTSPKANKEILDEAYVMAGVGSPLYVSRLLGICLTSTVQLV
TQLMPYGCLLDHVRENRLGSLQDLLNWCMIKAGMSYLEDVRLVHRDLAARNVLVKSPNHVKITDFGLA
RLLDIDETEHADGGKVPIKWMALESILRRRFTHQSDVWSYGVTVWELMTFGAKPYDGIPAREIPDLLEKGE
RLPQPPICTIDVYMIMVKCWMIDSECRPRFRELVSFESRMARDPQRFVVIQNEGLGPASPLDSTFYRSLLEDD
MGDLVDAEEYLVPQQGFFCPDPAPGAGGMVHHRSSSTRSGGDLTLGLEPSEEEAPRSPLAPSEGAGSDV
FDGDLGMGAAGLQSLPHTDPSPLQRYSEDPTVPLPSETDGYVAPLTCSPQPEYVNQPDVRPQPPSPREGPLP

AARPAGATLERPKTLSPGKNGVVKDVFAFGGAVENPEYLTQGGGAAPQPHPPAFSPAFDNLYYWDQDPPE
 RGAPPSTFKGTPTAENPEYLGLDVPV

Related Products

EGFR:HER2:HER3:HER4	
H_EGFR Reporter Cell Line	H_HER2 HER4 Reporter HEK-293 Cell Line
Cynomolgus_EGFR CHO-K1 Cell Line	Cynomolgus_ERBB3(HER3) CHO-K1 Cell Line
Cynomolgus_ERBB3(HER3) HEK-293 Cell Line	Cynomolgus_HER2(ERBB2) CHO-K1 Cell Line
H_EGFR CHO-K1 Cell Line	H_EGFR HEK-293 Cell Line
H_EGFRvIII CHO-K1 Cell Line	H_ERBB3(HER3) CHO-K1 Cell Line
H_ERBB3(HER3) HEK-293 Cell Line	H_ERBB3(HER3) MC38 Cell Line
H_HER2 HER3 MC38 Cell Line	H_HER2 MCF-7 Cell Line
H_HER2(ERBB2) CHO-K1 Cell Line	H_HER2(ERBB2) CT26 Cell Line
H_HER2(ERBB2) LLC1 Cell Line	H_HER2(ERBB2) MC38 Cell Line
Mouse_HER3(ERBB3) CHO-K1 Cell Line	H_EGFR LGR5 CT26 Cell Line
H_EGFR LGR5 HEK-293 Cell Line	
Anti-EGFR hIgG1 Reference Antibody (Cetbio)	Anti-EGFR hIgG1 Reference Antibody (Necbio)
Anti-ERBB3(HER3) hIgG1 Reference Antibody(Patibio)	Anti-H_EGFR hIgG1 Antibody(Necitumumab)
Anti-H_ERBB3(HER3) hIgG1 Antibody(Barecetamab)	Anti-H_HER2 hIgG1 Antibody(Margetuximab)
Anti-HER2 hIgG1 Reference Antibody(Marbio)	Anti-HER2 hIgG1 Reference Antibody(Trasbio)
Biotinylated Cynomolgus EGFR Protein; His-Avi Tag	Biotinylated Human EGFR Protein; His-Avi Tag
Biotinylated Human HER3 Protein; His-Avi Tag	Cynomolgus HER2 Protein; His Tag
Human EGFR Protein; hFc Tag	Human EGFR Protein; His Tag
Human HER2 Protein; hFc Tag	Human HER2 Protein; His Tag
Human HER3 Protein; hFc Tag	Human HER3 Protein; His Tag
Mouse HER3 Protein; His Tag	

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