Genomeditech (Shanghai) Co.,Ltd.
Order: +86 021-68455258/50432826/50432825
Toll-free: +86 400 627 9288
Email: service@genomeditech.com

Product Sheet

TCR Knockout Jurkat Cell Line(CD4-)

Catalog number: GM-C48309

Version 3.3.1.260730

Description	TCR Knockout Jurkat Cell Line(CD4-) is a clonal stable cell line derived from Jurkat cells with a knockout of Human CD4 and Human TCRαβ.
Quantity	5E6 Cells per vial,1 mL
Product Format	1 vial of frozen cells
Shipping	Shipped on dry ice
Storage Conditions	Liquid nitrogen immediately upon receipt
Target	Human CD4 and Human TCRαβ
Gene ID/Uniprot ID	/
Host Cell	Jurkat
Recovery Medium	RPMI 1640+10% FBS+1% P.S
Growth medium	RPMI 1640+10% FBS+1% P.S+400 μg/mL G418+200 μg/mL Hygromycin
Note	None
Freezing Medium	90% FBS+10% DMSO
Growth properties	Suspension
Growth Conditions	37°C, 5% CO_2
Mycoplasma Testing	The cell line has been screened to confirm the absence of Mycoplasma species.
Safety considerations	Biosafety Level 2
Note	It is recommended to expand the cell culture and store a minimum of 10 vials at an early passage for potential future use.

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Materials

Reagent	Manufacturer/Catalogue No.
RPMI 1640	gibco/C11875500BT
Fetal Bovine Serum	ExCell/FSP500
Pen/Strep	Thermo/15140-122
G418	Genomeditech/GM-040402
Hygromycin	Genomeditech/GM-040403
Anti-H_CD4 hIgG1 Antibody(Tregalizumab)	Genomeditech/GM-28752AB
FITC anti-human TCR α/β Antibody	Biolegend/306706

Figures

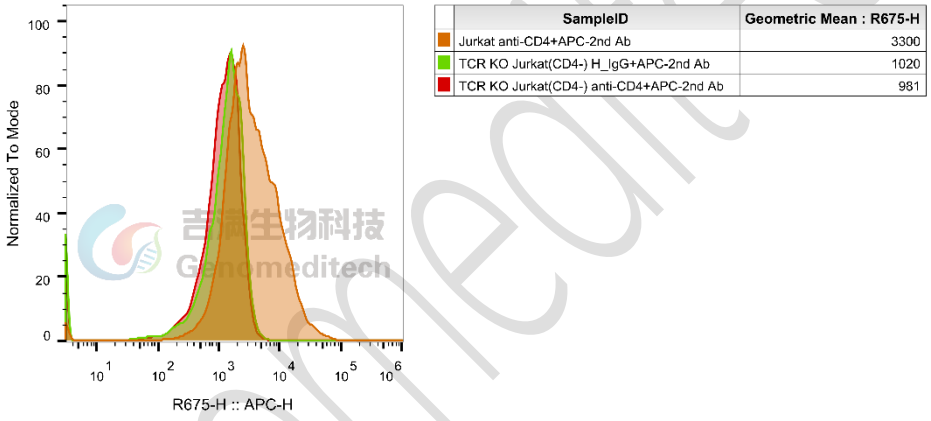

Figure 1 | TCR Knockout Jurkat Cell Line(CD4-) (Cat. GM-C48309) was determined by flow cytometry using Anti-H_CD4 hIgG1 Antibody(Tregalizumab) (Cat. GM-28752AB).

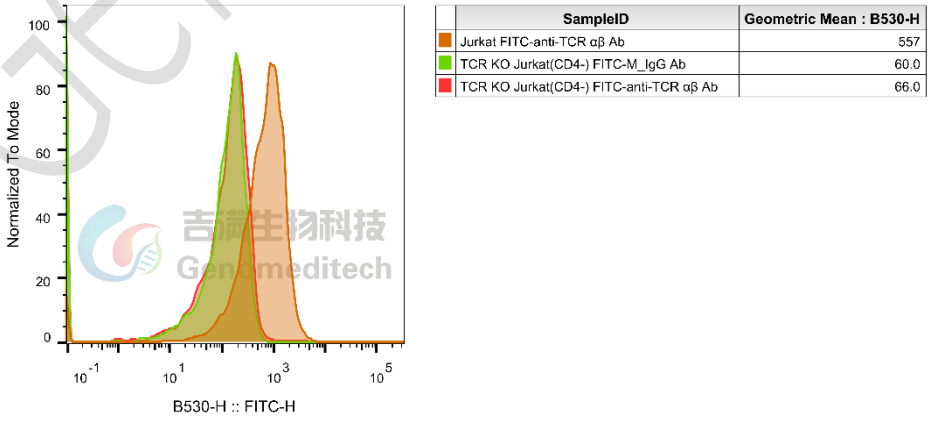

Figure 2 | TCR Knockout Jurkat Cell Line(CD4-) (Cat. GM-C48309) was determined by flow cytometry using FITC anti-human TCR α/β Antibody(Biolegend/306706).

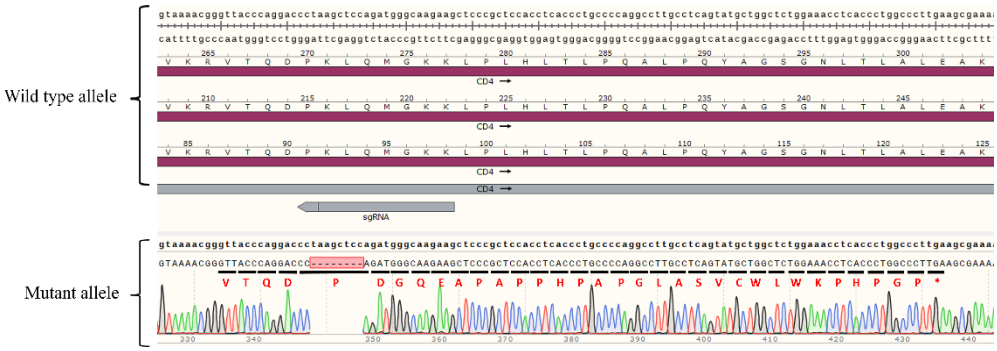

Figure 3 | The Sanger sequencing of the TCR Knockout Jurkat Cell Line showed successful knockout of CD4.

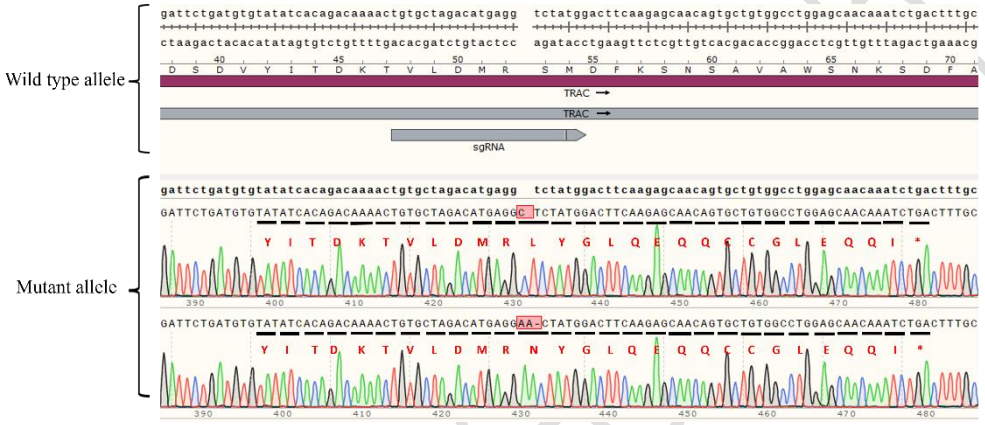

Figure 4 | The Sanger sequencing of the TCR Knockout Jurkat Cell Line showed successful knockout of TRAC.

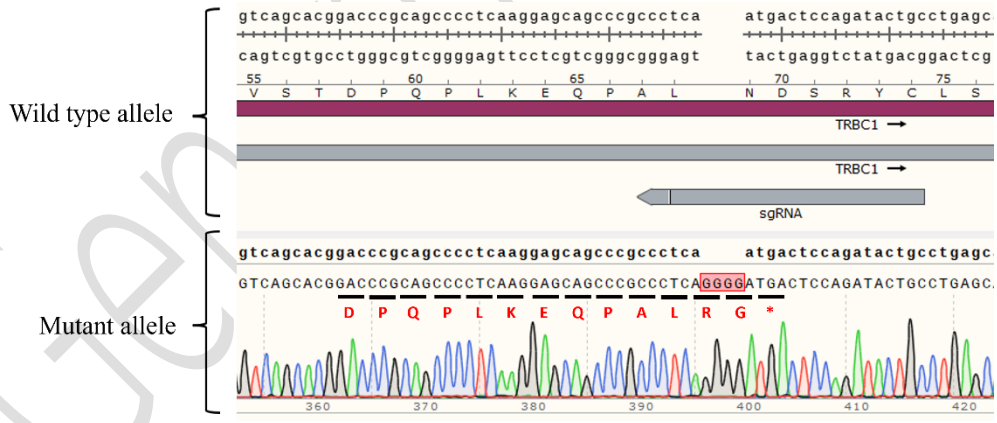

Figure 5 | The Sanger sequencing of the TCR Knockout Jurkat Cell Line showed successful knockout of TRBC.

Cell Recovery

Recovery Medium: RPMI 1640+10% FBS+1% P.S

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

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a) Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 - 3 minutes).
b) Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
c) Transfer the vial contents to a centrifuge tube containing 5.0 mL complete culture medium. And spin at approximately 176 x g for 5 minutes. Discard supernatant.
d) Resuspend cell pellet with the recommended complete medium. And dispense the suspension into 1 - 2 T-25 culture flasks.
e) Incubate the culture at 37°C in a suitable incubator. A 5% CO_2 in air atmosphere is recommended if using the medium described on this product sheet.

Cell Freezing

Freezing Medium: 90% FBS+10% DMSO

a) Centrifuge at 176 x g for 3 minutes to collect cells.
b) Resuspend the cells in pre-cooled freezing medium and adjust the cell density to 5E6 cells/mL.
c) Aliquot 1 mL into each vial.
d) Place the vial in a controlled-rate freezing container and store at -80°C for at least 1 day, then transfer to liquid nitrogen as soon as possible.

Cell passage

Growth medium: RPMI 1640+10% FBS+1% P.S+400 μg/mL G418+200 μg/mL Hygromycin

Approximately 48-72 hours after the initial thawing, the cells can be passaged for the first time. After this initial passage, the culture medium can be adjusted to growth medium supplemented with antibiotics. If cells are not passaged within 48 hours, it is recommended to add some fresh recovery medium and place the flask horizontally.

a) When the cell density reaches 1.5 - 2E6 cells/mL, subculture the cells. Do not allow the cell density to exceed 2E6 cells/mL.
b) It is recommended to use T-25 flasks for subculturing.
c) These cells are suspension cells, and it is recommended to use the "half-medium change" method to maintain optimal cell conditions during passaging.
d) During passaging, you can directly add fresh growth medium to the culture flask, gently pipette to resuspend the cells, and then transfer the cell suspension to a new T-25 flask for continued culture.

Subcultivation Ratio: Maintain cultures at a cell concentraion between 3E5 and 1E6 viable cells/mL.

Medium Renewal: Every 2 to 3 days

Notes

a) These cells are sensitive to density, so please ensure that the cell density is maintained within an appropriate range during culture and subculturing.

b) During the first passage, pay attention to the nutrient supply; if not subculturing, make sure to add fresh recovery medium every other day as needed.

Related Products

BTN3:Vγ9Vδ2	
TCR Knockout Jurkat Cell Line	H_Vγ9Vδ2(G115) Reporter Jurkat(TCRαβ KO) Cell Line
H_Vγ9Vδ2(MOP) Reporter Jurkat(TCRαβ KO) Cell Line	H_BTN2A1 HEK-293 Cell Line
H_BTN3A1 HEK-293 Cell Line	
Anti-BTN2A1 mIgG1 Antibody(mAb 7.48)	Anti-BTN3A1 hIgG1 Antibody(mAb1)
Anti-H_BTN3A1 hIgG1 Antibody(hu103.2)	Anti-Vγ9Vδ2 TCR hIgG1 Antibody
Human BTN3A1 Protein; His Tag	Human BTN3A2 Protein; His Tag
Human BTN3A3 Protein; His Tag	

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