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Product Sheet

Luciferase A375 Cell Line

Catalog number: GM-C45982

Version 4.0.26.1029

Specifications

Description	Luciferase A375 Cell Line is a clonal stable A375 cell line that constitutively expresses the Luciferase gene,constructed using lentiviral technology.
Quantity	5E6 Cells per vial,1 mL
Product Format	1 vials of frozen cells
Shipping	Shipped on dry ice
Storage Conditions	Liquid nitrogen immediately upon receipt
Target	Luciferase
Gene ID/Uniprot ID	无
Host Cell	A375 Cell Line
Recovery Medium	DMEM+10% FBS+1% P.S
Growth medium	DMEM+10% FBS+1% P.S+0.5 μg/mL Puromycin
Note	None
Freezing Medium	90% FBS+10% DMSO
Growth properties	Adherent
Growth Conditions	37°C, 5% CO_2
Mycoplasma Testing	The cell line has been screened to confirm the absence of Mycoplasma species.
Safety considerations	Biosafety Level 2
Note	It is recommended to expand the cell culture and store a minimum of 10 vials at an early passage for potential future use.

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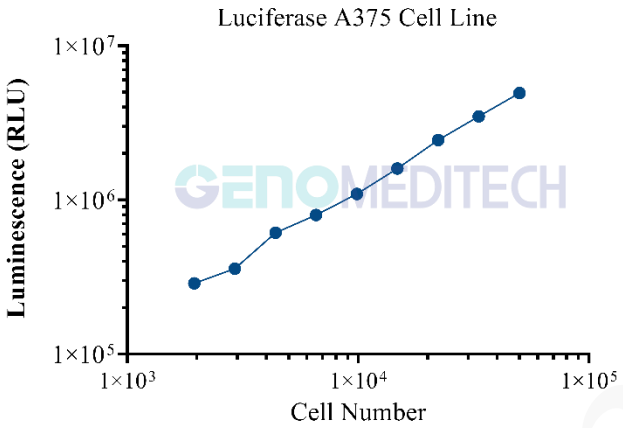
Materials

Reagent	Manufacturer/Catalogue No.
DMEM	Gibco/C11995500BT
Fetal Bovine Serum	ExCell/FSP500
Pen/Strep	Thermo/15140-122
Puromycin	Genomeditech/GM-040401
GMOne-Step 2.0 Luciferase Reporter Gene Assay Kit	Genomeditech/GM-040513
Countess 3 Automated Cell Counter	ThermoFisher/A49891

Data

Expression



Correlation between the number of cells and bioluminescence values. Serial dilutions of Luciferase A375 Cell Line (Cat. GM-C45982) (96-well format). The firefly luciferase activity was measured using the GMOne-Step 2.0 Luciferase Reporter Gene Assay Kit (Cat. GM-040513).

Cell Recovery

Recovery Medium: DMEM+10% FBS+1% P.S

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

a) Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 - 3 minutes).
b) Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
c) Transfer the vial contents to a centrifuge tube containing 5.0 mL complete culture medium. And spin at approximately 176 x g for 5 minutes. Discard supernatant.
d) Resuspend cell pellet with the recommended complete medium. And dispense the suspension into 1 - 2 T-25 culture flasks.
e) Incubate the culture at 37°C in a suitable incubator. A 5% CO_2 in air atmosphere is recommended if using the medium described on this product sheet.

Cell Freezing

Freezing Medium: 90% FBS+10% DMSO

a) Centrifuge at 176 x g for 3 minutes to collect cells.
b) Resuspend the cells in pre-cooled freezing medium and adjust the cell density to 5E6 cells/mL.
c) Aliquot 1 mL into each vial.
d) Place the vial in a controlled-rate freezing container and store at -80°C for at least 1 day, then transfer to liquid nitrogen as soon as possible.

Cell passage

Growth medium: DMEM+10% FBS+1% P.S+0.5 μg/mL Puromycin

For the first 1 to 2 passages post-resuscitation, use the recovery medium. Once the cells have stabilized, switch to a growth medium.

a) Remove and discard culture medium.
b) Briefly rinse the cell layer with PBS to remove all traces of serum that contains trypsin inhibitor.
c) Add 1.0 mL of 0.25% (w/v) Trypsin-EDTA solution to dish and observe cells under an inverted microscope until cell layer is dispersed (usually within 3 minutes at 37°C).
d) Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.
e) Add 2.0 mL of growth medium to mix well and aspirate cells by gently pipetting.
f) After centrifugation, resuspend the pellet and add appropriate aliquots of the cell suspension to new culture vessels.
g) Incubate cultures at 37°C.

Subcultivation Ratio: A subcultivation ratio of 1:2 - 1:5 is recommended

Medium Renewal: Every 2 to 3 days

Notes

a) FBS should be heat-inactivated at 56°C for 30 minutes to inactivate complement and certain viruses, with minimal impact on most growth factor and cytokine activities.

Related Products

Labeled Cells	
Luciferase-GFP MCF-7 Cell Line	Luciferase A498 Cell Line
Luciferase B16-F10 Cell Line	Luciferase Daudi Cell Line
Luciferase HEK-293 Cell Line	Luciferase HL-60 Cell Line
Luciferase Jurkat Cell Line	Luciferase MIA PaCa-2 Cell Line
Luciferase MM.1R Cell Line	Luciferase NCI-H929 Cell Line
Luciferase OVCAR3 Cell Line	Luciferase RAMOS Cell Line
Luciferase U-937 Cell Line	Luciferase-ZsGreen1 K562 Cell Line
Luciferase-ZsGreen1 Raji Cell Line	Luciferase 22RV1 Cell Line
Luciferase LNCaP Cell Line	
D-Luciferin, Potassium Salt	D-Luciferin, Sodium Salt

License Agreement

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