Genomeditech (Shanghai) Co.,Ltd.
Order: +86 021-68455258/50432826/50432825
Toll-free: +86 400 627 9288
Email: service@genomeditech.com

Product Sheet

H_STEAP1 KO LNCaP Cell Line

Catalog number: GM-C45886

Version 3.3.1.260624

Description	H_STEAP1 KO LNCaP Cell Line is a clonal stable cell line derived from LNCaP cells with a knockout of human STEAP1.
Quantity	5E6 Cells per vial,1 mL
Product Format	1 vial of frozen cells
Shipping	Shipped on dry ice
Storage Conditions	Liquid nitrogen immediately upon receipt
Target	H_STEAP1
Gene ID/Uniprot ID	/
Host Cell	LNCap
Recovery Medium	RPMI 1640+15% FBS+1% P.S
Growth medium	RPMI 1640+15% FBS+1% P.S
Note	None
Freezing Medium	90% FBS+10% DMSO
Growth properties	Adherent
Growth Conditions	37°C, 5% CO_2
Mycoplasma Testing	The cell line has been screened to confirm the absence of Mycoplasma species.
Safety considerations	Biosafety Level 1
Note	It is recommended to expand the cell culture and store a minimum of 10 vials at an early passage for potential future use.

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Materials

Reagent	Manufacturer/Catalogue No.
Pen/Strep	Thermo/15140-122
Fetal Bovine Serum	Gibco/A5669701
RPMI 1640	gibco/C11875500BT
Anti-H_STEAP1 hIgG1 Antibody(Vandortuzumab)	Genomeditech/GM-33026AB

Figures

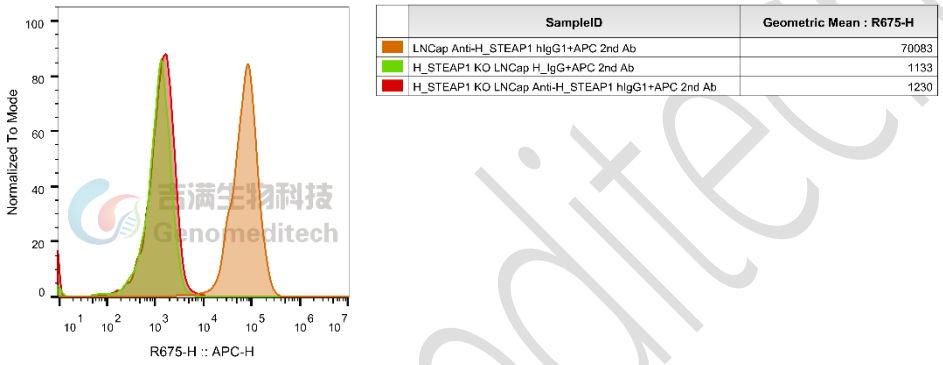

Figure 1 | H_STEAP1 KO LNCaP Cell Line (Cat. GM-C45886) was determined by flow cytometry using Anti-H_STEAP1 hIgG1 Antibody(Vandortuzumab) (Genomeditech/GM-33026AB).

Figure 2 | The Sanger sequencing of the H_STEAP1 KO LNCaP Cell Line showed successful knockout of STEAP1.

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Cell Recovery

Recovery Medium: RPMI 1640+15% FBS+1% P.S

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

a) Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 - 3 minutes).

b) Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.

c) Transfer the vial contents to a centrifuge tube containing 5.0 mL complete culture medium. And spin at approximately 176 x g for 5 minutes. Discard supernatant.

d) Resuspend cell pellet with the recommended complete medium. And dispense the suspension into an appropriate culture flask and initially place the flask in an upright position after thawing.

e) Incubate the culture at 37°C in a suitable incubator. A 5% CO_2 in air atmosphere is recommended if using the medium described on this product sheet.

Cell Freezing

Freezing Medium: 90% FBS+10% DMSO

a) Centrifuge at 176 x g for 3 minutes to collect cells.

b) Resuspend the cells in pre-cooled freezing medium and adjust the cell density to 5E6 cells/mL.

c) Aliquot 1 mL into each vial.

d) Place the vial in a controlled-rate freezing container and store at -80°C for at least 1 day, then transfer to liquid nitrogen as soon as possible.

Cell passage

Growth medium: RPMI 1640+15% FBS+1% P.S

For the first 1 to 2 passages post-resuscitation, use the recovery medium. Once the cells have stabilized, switch to a growth medium.

a) Subculturing is necessary when the cell density reaches 80%. It is recommended to perform subculturing at a ratio of 1:2 every 4-5 days. Ensure that the density does not exceed 90%, as overcrowding can lead to reduced viability due to compression.

b) Remove and discard culture medium.

c) Briefly rinse the cell layer with PBS to remove all traces of serum that contains trypsin inhibitor.

d) Add 1.0 mL of 0.25% (w/v) Trypsin-EDTA solution to dish and observe cells under an inverted microscope until cell layer is dispersed (usually within 3 to 5 minutes at 37°C).

e) Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.

f) Add 2.0 mL of growth medium to mix well and aspirate cells by gently pipetting.

g) After centrifugation, resuspend the pellet and add appropriate aliquots of the cell suspension to new culture vessels.

h) Incubate cultures at 37°C.

Subcultivation Ratio: A subcultivation ratio of 1:2 is recommended

Medium Renewal: Every 4 to 5 days

Notes

a) Cells immediately after thawing typically contain a certain proportion of dead cells, which is a normal phenomenon. With appropriate adjustment and recovery, cell viability generally improves markedly. Once the culture stabilizes, the proportion of dead cells decreases after subsequent passages, and the cell growth rate becomes more consistent.

b) Standard tissue culture (TC)-treated culture flasks or dishes may not provide optimal cell adhesion, which can increase the difficulty of cell culture. It is therefore recommended to pre-coat the culture surface with poly-L-lysine solution to enhance cell attachment.

c) FBS should be heat-inactivated at 56 °C for 30 minutes in a water bath. This process inactivates complement and some viruses, while generally having minimal impact on the activity of most growth factors and cytokines.

Related Products

STEAP1	
Cynomolgus_STEAP1 CHO-K1 Cell Line	H_STEAP1 CHO-K1 Cell Line
H_STEAP1 HEK-293 Cell Line	H_STEAP1 RM-1 Cell Line(Low Expression)
Mouse_STEAP1 CHO-K1 Cell Line	Rat_STEAP1 CHO-K1 Cell Line
Anti-H_STEAP1 hIgG1 Antibody(Vandortuzumab)	Anti-STEAP1 hIgG1 Reference Antibody (Vandbio)
STEAP2	
H_STEAP2 HEK-293 Cell Line	H_STEAP2(ECD) CHO-K1 Cell Line
H_STEAP2(ECD) HEK-293 Cell Line	H_STEAP2(ECD2) CHO-K1 Cell Line
H_STEAP2(ECD2) HEK-293 Cell Line	Mouse_STEAP2(ECD) HEK-293 Cell Line
Anti-STEAP2 hIgG1 Reference Antibody (AZD0754)	
STEAP3	
H_STEAP3-eGFP HEK-293 Cell Line	
STEAP4	
H_STEAP4-eGFP HEK-293 Cell Line	

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