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Product Sheet

H_CD38 HEK-293 Cell Line

Catalog number: GM-C49011

Version 4.0.26.1012

Specifications

Description	H_CD38 HEK-293 Cell Line is a clonal stable HEK-293 cell line that constitutively expresses the human CD38 gene, constructed using lentiviral technology.
Quantity	5E6 Cells per vial,1 mL
Product Format	1 vial of frozen cells
Shipping	Shipped on dry ice
Storage Conditions	Liquid nitrogen immediately upon receipt
Target	H_CD38
Gene ID/Uniprot ID	P28907-1 2004-11-23 v2
Host Cell	HEK-293 Cell Line
Recovery Medium	DMEM+10% FBS+1% P.S
Growth medium	DMEM+10% FBS+1% P.S+0.75 μg/mL Puromycin
Note	None
Freezing Medium	90% FBS+10% DMSO
Growth properties	Adherent
Growth Conditions	37°C, 5% CO_2
Mycoplasma Testing	The cell line has been screened to confirm the absence of Mycoplasma species.
Safety considerations	Biosafety Level 2
Note	It is recommended to expand the cell culture and store a minimum of 10 vials at an early passage for potential future use.

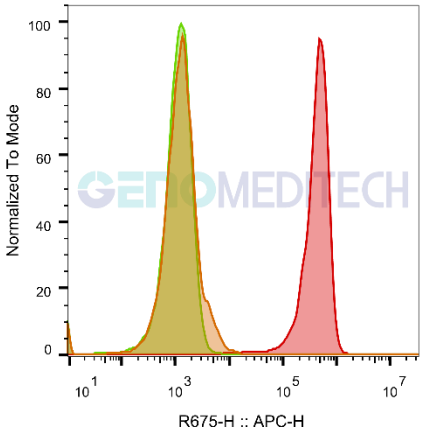
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Materials

Reagent	Manufacturer/Catalogue No.
DMEM	Gibco/C11995500BT
Fetal Bovine Serum	ExCell/FSP500
Pen/Strep	Thermo/15140-122
Puromycin	Genomeditech/GM-040401
Anti-CD38 hIgG1 Antibody(Daratumumab)	Genomeditech/GM-27202AB

Data

Expression



	SampleID	Geometric Mean : R675-H
	HEK-293 Anti-CD38+APC 2nd Ab	1211
	HEK-293 H_CD38 H_IgG+APC 2nd Ab	1047
	HEK-293 H_CD38 Anti-CD38+APC 2nd Ab	392808

H_CD38 HEK-293 Cell Line (Cat. GM-C49011) was determined by flow cytometry using Anti-CD38 hIgG1 Antibody(Daratumumab) (Cat. GM-27202AB).

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Cell Recovery

Recovery Medium: DMEM+10% FBS+1% P.S

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

a) Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 - 3 minutes).
b) Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
c) Transfer the vial contents to a centrifuge tube containing 5.0 mL complete culture medium. And spin at approximately 176 x g for 5 minutes. Discard supernatant.
d) Adjust the viable cell density to 2–3E5 cells/mL by supplementing with recovery medium. Based on the total volume of the cell suspension, seed the cells into appropriate culture vessels.
e) Incubate the culture at 37°C in a suitable incubator. A 5% CO_2 in air atmosphere is recommended if using the medium described on this product sheet.

Cell Freezing

Freezing Medium: 90% FBS+10% DMSO

a) Centrifuge at 176 x g for 3 minutes to collect cells.
b) Resuspend the cells in pre-cooled freezing medium and adjust the cell density to 5E6 cells/mL.
c) Aliquot 1 mL into each vial.
d) Place the vial in a controlled-rate freezing container and store at -80°C for at least 1 day, then transfer to liquid nitrogen as soon as possible.

Cell passage

Growth medium: DMEM+10% FBS+1% P.S+0.75 μg/mL Puromycin

For the first 1 to 2 passages post-resuscitation, use the recovery medium. Once the cells have stabilized, switch to a growth medium.

a) Remove and discard culture medium.
b) Briefly rinse the cell layer with PBS to remove all traces of serum that contains trypsin inhibitor.
c) Add 1.0 mL of 0.25% (w/v) Trypsin-EDTA solution to dish and observe cells under an inverted microscope until cell layer is dispersed (usually within 30 to 60 seconds at 37°C).
d) Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.
e) Add 2.0 mL of growth medium to mix well and aspirate cells by gently pipetting.
f) After centrifugation, resuspend the pellet and add appropriate aliquots of the cell suspension to new culture vessels.
g) Incubate cultures at 37°C.

Subcultivation Ratio: A subcultivation ratio of 1:2 - 1:3 is recommended

Medium Renewal: Every 2 to 3 days

Notes

a) A certain proportion of dead cells is commonly observed immediately after thawing, which is a normal phenomenon. The situation will improve markedly following an adaptation period; once the culture has stabilized, the number of dead cells after passaging decreases and the cell growth rate becomes consistent. During the first 1–3 days post-thaw, the cells may show poor or no attachment. If this occurs, the serum concentration can be increased by an additional 5% and the observation period extended accordingly. Generally, cells begin to attach on day 4, and the first passage can be performed on day 5. After a single passage at a 1:2 split ratio, cell growth gradually returns to normal, reaching over 80% confluence within 2–3 days.

b) It is important to maintain the cell density below 80%, as exceeding this threshold can lead to decreased cell viability and metabolic activity due to overcrowding.

c) FBS requires heat inactivation at 56°C for 30 minutes, which can inactivate complement and some viruses, but does not significantly affect the activity of most growth factors and cytokines.

Sequence

H_CD38 P28907-1 2004-11-23 v2

MANCEFSPVSGDKPCCRLSRRAQLCLGVSILVLILVVVLAVVVPRWRQQWSGPGTTKRFPETVLARCVKYTEIHP
EMRHVDCQSVWDAFKGAFISKHPCNITEEDYQPLMKLGTQTVPCNKILLWSRIKDLAHQFTQVQRDMFTLEDTL
LGYLADDLTWCGEFNTSKINYQSCPDWRKDCSNNPVSVFWKTVSRRFAEAACDVVHVMLNGSRSKIFDKNSTF
GSVEVHNLQPEKVQTLEAWVIHGGREDSRDLCQDPTIKELESIISKRNIQFSCKNIYRPDKFLQCVKNPEDSSCTSE
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Related Products

CD38	
Cynomolgus_CD38 CHO-K1 Cell Line	Cynomolgus_CD38 HEK-293 Cell Line
H_CD38 CHO-K1 Cell Line	
Anti-CD38 hIgG1 Antibody(Daratumumab)	

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