Genomeditech (Shanghai) Co.,Ltd.
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Product Sheet

H_CDH17 SW480 Cell Line

Catalog number: GM-C31741

Version 3.3.1.251226

Description	H_CDH17 SW480 Cell Line is a clonal stable SW480 cell line that constitutively expresses the human CDH17 gene, constructed using lentiviral technology.
Quantity	5E6 Cells per vial,1 mL
Product Format	1 vial of frozen cells
Shipping	Shipped on dry ice
Storage Conditions	Liquid nitrogen immediately upon receipt

Target	Human_CDH17
Gene ID/Uniprot ID	Q12864(AA Met 1 - Ile 808)
Host Cell	SW480

Recovery Medium	RPMI 1640+10% FBS+1% P.S
Growth medium	RPMI 1640+10% FBS+1% P.S+5 μg/mL Blasticidin+0.5 μg/mL Puromycin
Note	None
Freezing Medium	90% FBS+10% DMSO
Growth properties	Adherent
Growth Conditions	37°C, 5% CO_2

Mycoplasma Testing	The cell line has been screened to confirm the absence of Mycoplasma species.
Safety considerations	Biosafety Level 2
Note	It is recommended to expand the cell culture and store a minimum of 10 vials at an early passage for potential future use.

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Materials

Reagent	Manufacturer/Catalogue No.
RPMI 1640	VivaCell/C3010-0500
Fetal Bovine Serum	ExCell/FSP500
Pen/Strep	Thermo/15140-122
Blasticidin	Genomeditech/GM-040404
Puromycin	Genomeditech/GM-040401
Anti-CDH17 hIgG1 Antibody(BI-905711)	Genomeditech/GM-52672AB

Figures

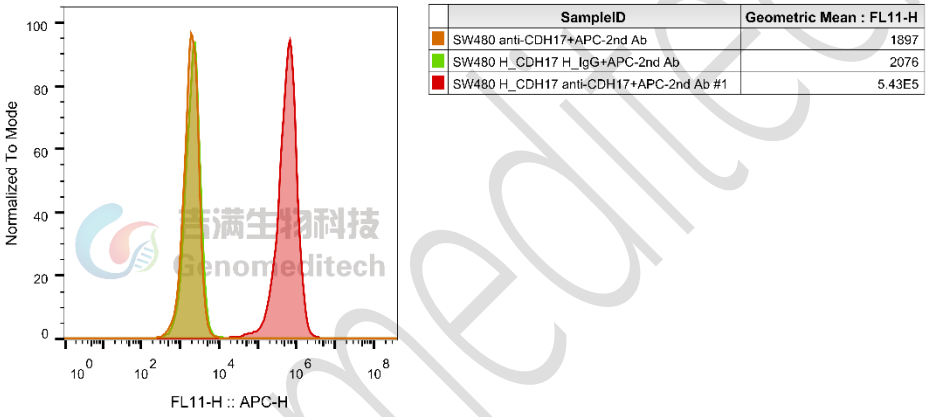

Figure 1 | H_CDH17 SW480 Cell Line (Cat. GM-C31741) was determined by flow cytometry using Anti-CDH17 hIgG1 Antibody(BI-905711) (Cat. GM-52672AB).

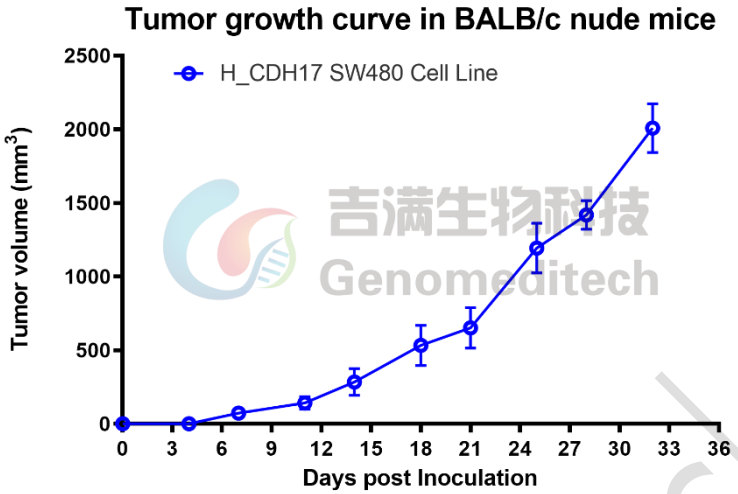

Figure 2 | Tumor growth curves of H_CDH17 SW480 in BALB/c nude mice. H_CDH17 SW480 cells (1×10^6 per mouse) were subcutaneously inoculated into BALB/c nude mice (female, 8 weeks old, n = 3). Tumor volume was measured twice per week and is presented as mean ± SEM.

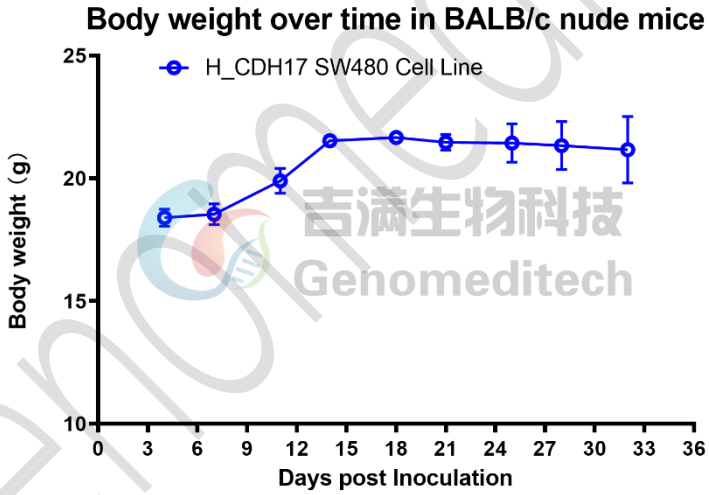

Figure 3 | Body weight changes after implantation of H_CDH17 SW480 in BALB/c nude mice. Under the same conditions, body weight was measured twice per week and is presented as mean ± SEM.

Cell Recovery

Recovery Medium: RPMI 1640+10% FBS+1% P.S

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

a) Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 - 3 minutes).
b) Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
c) Transfer the vial contents to a centrifuge tube containing 5.0 mL complete culture medium and spin at approximately 176 x g for 5 minutes. Discard supernatant.
d) Resuspend cell pellet with the recommended recovery medium. And dispense into appropriate culture dishes.
e) Incubate the culture at 37°C in a suitable incubator. A 5% CO_2 in air atmosphere is recommended if using the medium described on this product sheet.

Cell Freezing

Freezing Medium: 90% FBS+10% DMSO

a) Centrifuge at 176 x g for 3 minutes to collect cells.
b) Resuspend the cells in pre-cooled freezing medium and adjust the cell density to 5E6 cells/mL.
c) Aliquot 1 mL into each vial.
d) Place the vial in a controlled-rate freezing container and store at -80°C for at least 1 day, then transfer to liquid nitrogen as soon as possible.

Cell passage

Growth medium: RPMI 1640+10% FBS+1% P.S+5 μg/mL Blasticidin+0.5 μg/mL Puromycin

For the first 1 to 2 passages post-resuscitation, use the recovery medium. Once the cells have stabilized, switch to a growth medium.

a) Remove and discard culture medium.
b) Briefly rinse the cell layer with PBS to remove all traces of serum that contains trypsin inhibitor.
c) Add 1.0 mL of 0.25% (w/v) Trypsin-EDTA solution to dish and observe cells under an inverted microscope until cell layer is dispersed (usually within 2 to 3 minutes at 37°C).
d) Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.
e) Add 2.0 mL of growth medium to mix well and aspirate cells by gently pipetting.
f) After centrifugation, resuspend the pellet and add appropriate aliquots of the cell suspension to new culture vessels.
g) Incubate cultures at 37°C.

Subcultivation Ratio: A subcultivation ratio of 1:2 - 1:3 is recommended

Medium Renewal: Every 2 to 3 days

Notes

a) After the stabilization of the cell condition, there will be fewer dead cells post-passage, the cell growth rate will tend to stabilize, cell morphology will become uniform, and the cells will appear robust.

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Sequence

CDH17 Q12864(ΔICD)

MILQAHLHSLCLLMLYLATGYGQEGKFSGPLKPMTFSIYEGQEPSQIIFQFKANPPAVTFELTGETDNIFVIERE
GLLYYNRALDRETRSTHNLQVAALDANGIIVEGPVPITIKVKDINDNRPTFLQSKYEGSVRQNSRPGKPFLYV
NATDLDDPATPNGQLYYQIVIQLPMINNVMYFQINNKTGAISLTREGSQELNPAKNPSYNLVISVKDMGGQSE
NSFSDTTSVDIIVTENIWKAPKPVEMVENSTDPHPIKITQVRWNDPGAQYSLVDKEKLPRFPFSIDQEGDIYVT
QPLDREEKDAYVFYAVAKDEYGKPLSYPLEIHVKVKDINDNPPTCPSPVTVFEVQENERLGNSIGTLTAHDRD
EENTANSFLNYRIVEQTPKLPMDGLFLIQTYAGMLQLAKQSLKKQDTPQYNLTIEVSDKDFKTLCFVQINVIDI
NDQIPIFEKSDYGNLTLAEDTNIGSTILTIQATDADEPFTGSSKILYHIIKGDSEGRLGVDTDPHTNTGYVIIKKP
LDFETAAVSNIVFKAENPEPLVFGVKYNASSFAKFTLIVTDVNEAPQFSQHVFQAKVSEDVAIGTKVGNVTAK
DPEGLDISYSLRGDTRGWLKIDHVTGEIFSVAPLDREAGSPYRVQVVATEVGGSSLSSVSEFHLILMDVNDNP
PRLAKDYTGLFFCHPLSAPGSLIFEATDDDQHLFRGPHFTFSLGSGSLQNDWEVSKINGTHARLSTRHTEFEER
EYVVLIRINDGGRPPLEGIVSLPVTFCSCVEGSCFRPAGHQTGIPTVGMAVGILLTTLLVIGIILAVVFI*

Related Products

CDH3	
Cynomolgus_CDH3 CHO-K1 Cell Line	H_CDH3 CHO-K1 Cell Line
H_CDH3 HEK-293 Cell Line	
Anti-H_CDH3 hIgG1 Antibody	
CDH6	
Cynomolgus_CDH6 CHO-K1 Cell Line	H_CDH6 CHO-K1 Cell Line
H_CDH6 HEK-293 Cell Line	
Anti-CDH6 hIgG1 Reference Antibody (Ralubio)	Anti-H_CDH6 hIgG1 Antibody(H01L02)
Cynomolgus CDH6 Protein; His Tag	Human CDH6 Protein; His Tag
CDH17	
Cynomolgus_CDH17 HEK-293 Cell Line	Cynomolgus_CDH17(XP_005563762.1) HEK-293 Cell Line
H_CDH17 CHO-K1 Cell Line	H_CDH17 CT26 Cell Line
H_CDH17 HCT116 Cell Line	H_CDH17 HEK-293 Cell Line
H_CDH17 LLC1 Cell Line	H_CDH17 MC38 Cell Line
H_CDH17 RKO Cell Line	H_CDH17(ΔEC1,Flag-EC2-7) HEK-293 Cell Line
H_CDH17(ΔEC1-2,Flag-EC3-7) HEK-293 Cell Line	H_CDH17(ΔEC1-3,Flag-EC4-7) HEK-293 Cell Line
H_CDH17(ΔEC1-4,Flag-EC5-7) HEK-293 Cell Line	H_CDH17(ΔEC1-5,Flag-EC6-7) HEK-293 Cell Line
H_CDH17(ΔEC1-6,Flag-EC7) HEK-293 Cell Line	Mouse_CDH17 HEK-293 Cell Line
Rat_CDH17 HEK-293 Cell Line	Rhesus_CDH17 HEK-293 Cell Line
Anti-CDH17 hIgG1 Antibody(BI-905711)	Anti-CDH17 hIgG1 Antibody(VHHl-28BB)
Anti-CDH17 hIgG1 Reference Antibody(BI-905711)	
Biotinylated Human CDH17 Protein; His-Avi Tag	Cynomolgus CDH17 Protein; His Tag
Human CDH17 Protein; His Tag	Mouse CDH17 Protein; His Tag

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