

# Product Sheet

## H\_FGF21 Reporter HEK-293 Cell Line

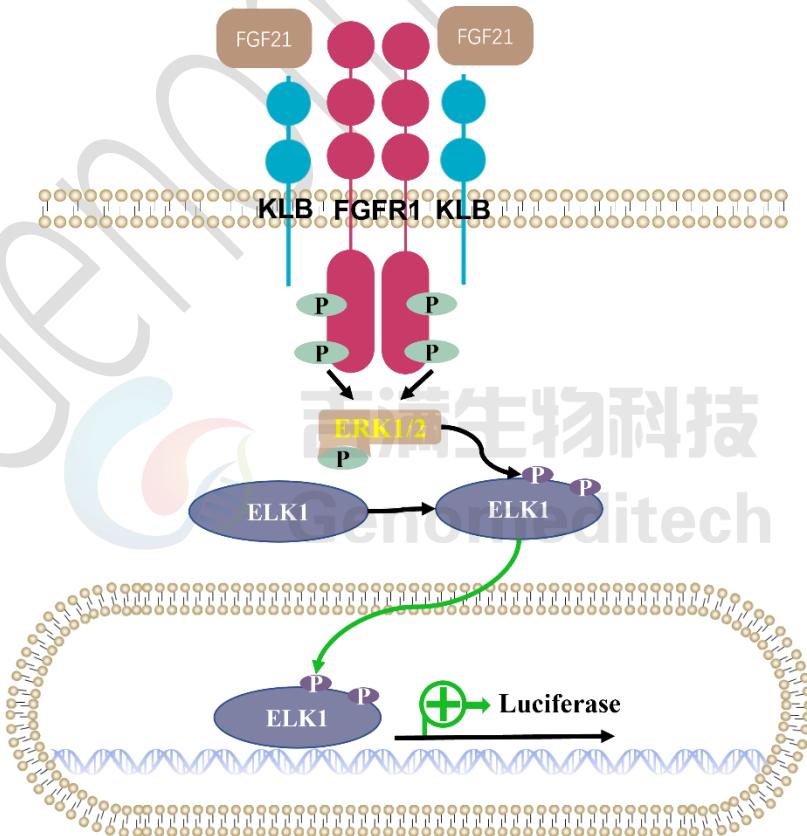
Catalog number: GM-C19834

Version 3.3.1.260109

FGF21 (Fibroblast Growth Factor 21) is a protein secreted by the liver, adipose tissue, and muscle, belonging to the FGF family. It is essential for energy metabolism, glucose homeostasis, and fatty acid oxidation, especially during hunger and metabolic stress. FGF21 regulates these processes by binding to its receptor FGFR and activating downstream signaling pathways, with secretion influenced by nutritional status, insulin levels, and inflammation.

The FGF21 signaling pathway is mediated by FGFR and the co-receptor  $\beta$ -klotho. Binding to FGFR activates downstream pathways like MAPK and PI3K/Akt, promoting fatty acid oxidation, glucose uptake, and insulin sensitivity. FGF21 also regulates gene expression in the liver and adipose tissue, impacting energy and lipid metabolism. Thus, FGF21 is a potential therapeutic target for metabolic syndrome and diabetes.

H\_FGF21 Reporter HEK-293 Cell Line is a clonal stable HEK-293 cell line constructed using lentiviral technology, constitutive expression of the Klotho $\beta$  and FGFR1c gene, along with signal-dependent expression of a luciferase reporter gene. When FGF21 binds to FGF21 Receptor Signaling Complex, it activates downstream signaling pathways, leading to the expression of luciferase. The luciferase activity measurement indicates the activation level of the signaling pathway and can thus be used to evaluate the in vitro effects of drugs related to FGF21.



## Specifications

<b>Quantity</b>	5E6 Cells per vial, 1 mL
<b>Product Format</b>	1 vial of frozen cells
<b>Shipping</b>	Shipped on dry ice
<b>Storage Conditions</b>	Liquid nitrogen immediately upon receipt
<b>Recovery Medium</b>	DMEM+10% FBS+1% P.S
<b>Growth medium</b>	DMEM+10% FBS+1% P.S+4 $\mu$ g/mL Blasticidin+400 $\mu$ g/mL G418+125 $\mu$ g/mL Hygromycin+0.75 $\mu$ g/mL Puromycin
<b>Note</b>	None
<b>Freezing Medium</b>	90% FBS+10% DMSO
<b>Growth properties</b>	Adherent
<b>Growth Conditions</b>	37°C, 5% CO <sub>2</sub>
<b>Mycoplasma Testing</b>	The cell line has been screened to confirm the absence of Mycoplasma species.
<b>Safety considerations</b>	Biosafety Level 2
<b>Note</b>	It is recommended to expand the cell culture and store a minimum of 10 vials at an early passage for potential future use.

## Materials

Reagent	Manufacturer/Catalogue No.
DMEM	Gibco/C11995500BT
Fetal Bovine Serum	ExCell/FSP500
Pen/Strep	Thermo/15140-122
Blasticidin	Genomeditech/ <a href="#">GM-040404</a>
G418	Genomeditech/ <a href="#">GM-040402</a>
Hygromycin	Genomeditech/ <a href="#">GM-040403</a>
Puromycin	Genomeditech/ <a href="#">GM-040401</a>
Recombinant Human FGF-21	R&D SYSTEMS/2539-FG/CF
Human FGF <sub>a</sub> (140AA)	Novoprotein/C049
Human FGF <sub>b</sub> (157AA)	Novoprotein/C046
Anti-Human CD331/FGFR1 Antibody (A08)	antibodysystem/FHC88110
RG7992 (anti-Klotho Beta)	Aladdin/Ab177889
GMOne-Step 2.0 Luciferase Reporter Gene Assay Kit	Genomeditech/ <a href="#">GM-040513</a>

## Figures

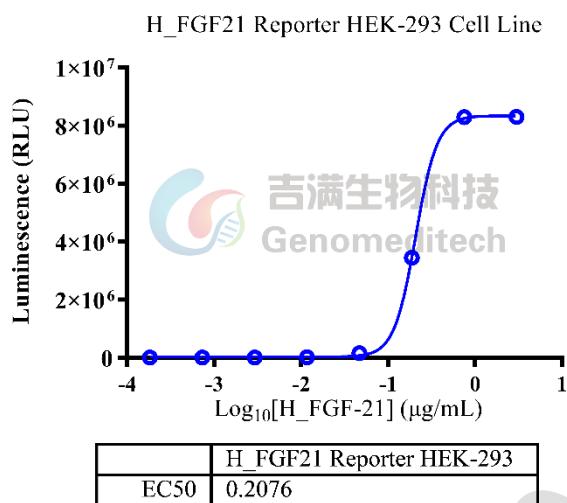


Figure 1 | Response to Recombinant Human FGF-21. The H\_FGF21 Reporter HEK-293 Cell Line (Cat. GM-C19834) at a concentration of 1.5E4 cells/well (96-well format) was stimulated with serial dilutions of Recombinant Human FGF-21 (R&D SYSTEMS/2539-FG/CF) in assay buffer (DMEM + 1% FBS + 1% P.S) for 16 hours. The firefly luciferase activity was measured using the Luciferase Reporter Assay Kit (Genomeditech). The maximum induction fold was approximately. The maximum induction fold was approximately [1602.8]. Data are shown by drug mass concentration.

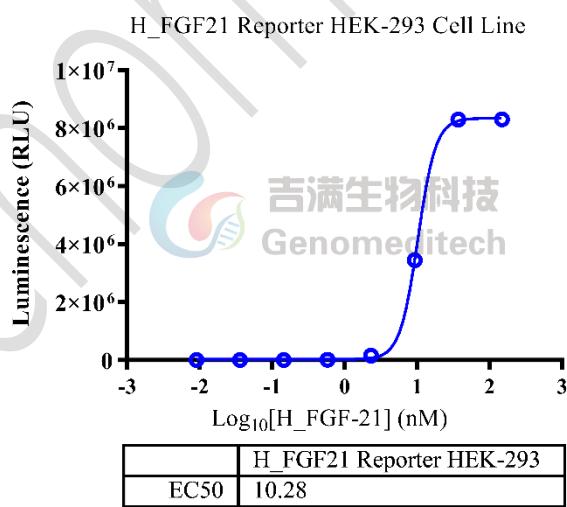


Figure 2 | Response to Recombinant Human FGF-21. The H\_FGF21 Reporter HEK-293 Cell Line (Cat. GM-C19834) at a concentration of 1.5E4 cells/well (96-well format) was stimulated with serial dilutions of Recombinant Human FGF-21 (R&D SYSTEMS/2539-FG/CF) in assay buffer (DMEM + 1% FBS + 1% P.S) for 16 hours. The firefly luciferase activity was measured using the Luciferase Reporter Assay Kit (Genomeditech). The maximum induction fold was approximately. The maximum induction fold was approximately [1602.8]. Data are shown by drug molar concentration.

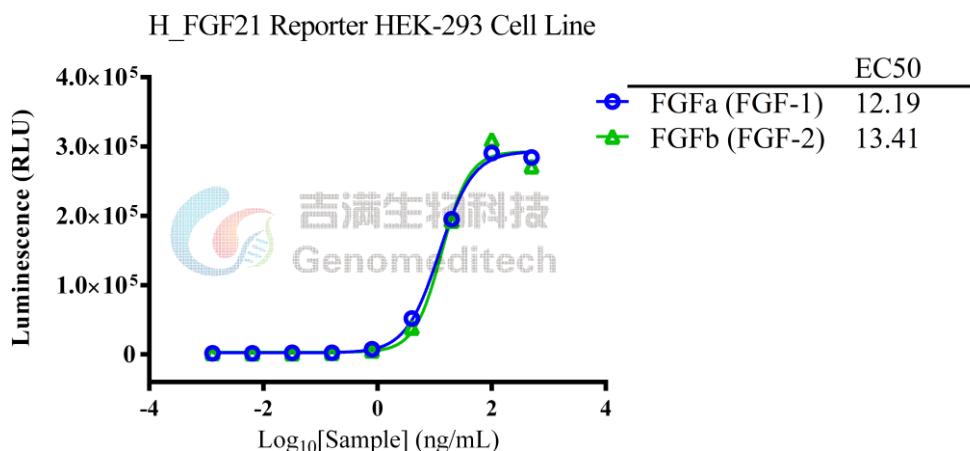


Figure 3 | Response to Human FGFa (140AA) and Human FGFb (157AA). The H\_FGF21 Reporter HEK-293 Cell Line (Cat. GM-C19834) at a concentration of 1.3E4 cells/well (96-well format) was stimulated with serial dilutions of Human FGFa (140AA) (Novoprotein/C049) and Human FGFb (157AA) (Novoprotein/C046) in assay buffer (DMEM + 1% FBS + 1% P.S) for 6 hours. The firefly luciferase activity was measured using the Luciferase Reporter Assay Kit (Genomeditech). The maximum induction fold was approximately. The maximum induction fold was approximately [125.1] and [165.0], respectively. Data are shown by drug mass concentration.

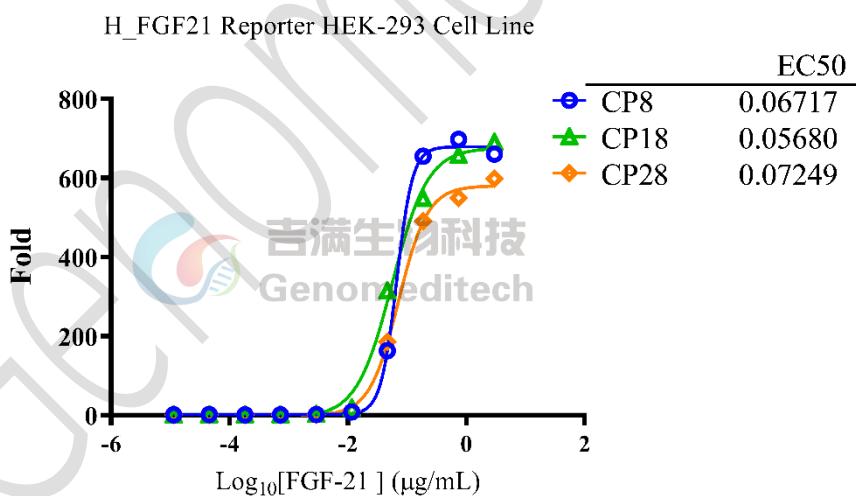


Figure 4 | The passage stability of response to Recombinant Human FGF-21 Protein. The passage 8, 18 and 28 of H\_FGF21 Reporter HEK-293 Cell Line (Cat. GM-C19834) at a concentration of 1.3E4 cells/well (96-well format) were stimulated with serial dilutions of Recombinant Human FGF-21 Protein (R&D SYSTEMS/2539-FG/CF) in assay buffer (DMEM + 1% FBS + 1% P.S) for 16 hours. The firefly luciferase activity was measured using the Luciferase Reporter Assay Kit (Genomeditech). The maximum induction fold was approximately. Data are shown by drug mass concentration.

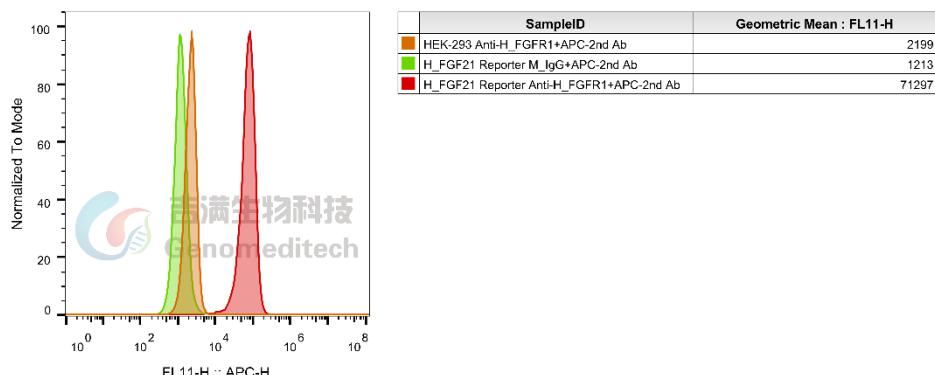


Figure 5 | H\_FGF21 Reporter HEK-293 Cell Line (Cat. GM-C19834) was determined by flow cytometry using Anti-Human CD331/FGFR1 Antibody (A08) (antibodysystem/FHC88110).

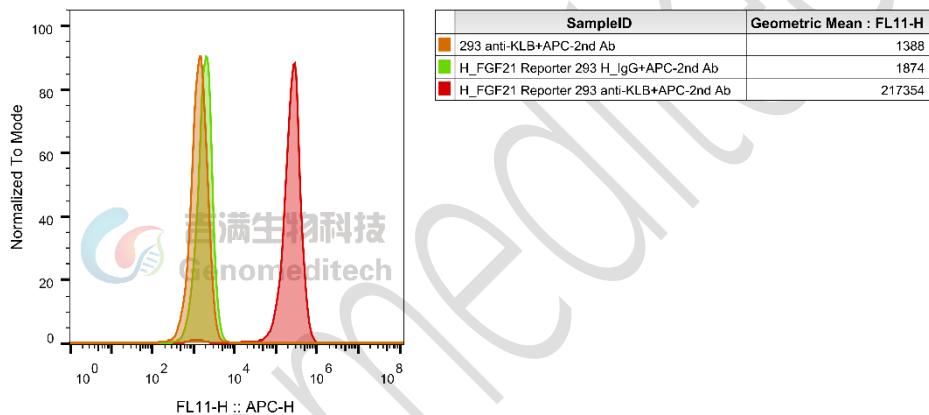


Figure 6 | H\_FGF21 Reporter HEK-293 Cell Line (Cat. GM-C19834) was determined by flow cytometry using RG7992 (anti-Klotho Beta) (Aladdin/Ab177889).

## Cell Recovery

Recovery Medium: DMEM+10% FBS+1% P.S

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

- Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 - 3 minutes).
- Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
- Transfer the vial contents to a centrifuge tube containing 5.0 mL complete culture medium and spin at approximately 176 x g for 5 minutes. Discard supernatant.

- d) Resuspend cell pellet with the recommended recovery medium. And dispense into appropriate culture dishes.
- e) Incubate the culture at 37°C in a suitable incubator. A 5% CO<sub>2</sub> in air atmosphere is recommended if using the medium described on this product sheet.

## Cell Freezing

Freezing Medium: 90% FBS+10% DMSO

- a) Centrifuge at 176 x g for 3 minutes to collect cells.
- b) Resuspend the cells in pre-cooled freezing medium and adjust the cell density to 5E6 cells/mL.
- c) Aliquot 1 mL into each vial.
- d) Place the vial in a controlled-rate freezing container and store at -80°C for at least 1 day, then transfer to liquid nitrogen as soon as possible.

## Cell passage

Growth medium: DMEM+10% FBS+1% P.S+4 µg/mL Blasticidin+400 µg/mL G418+125 µg/mL Hygromycin+0.75 µg/mL Puromycin

For the first 1 to 2 passages post-resuscitation, use the recovery medium. Once the cells have stabilized, switch to a growth medium.

- a) Subculturing is necessary when the cell density reaches 80%. It is recommended to perform subculturing at a ratio of 1:3 to 1:4 every 2-3 days. Ensure that the density does not exceed 80%, as overcrowding can lead to reduced viability due to compression.
- b) Remove and discard culture medium.
- c) Briefly rinse the cell layer with PBS to remove all traces of serum that contains trypsin inhibitor.
- d) Add 1.0 mL of 0.25% (w/v) Trypsin-EDTA solution to dish and observe cells under an inverted microscope until cell layer is dispersed (usually within 30 to 60 seconds at 37°C).
- e) Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.
- f) Add 2.0 mL of growth medium to mix well and aspirate cells by gently pipetting.
- g) After centrifugation, resuspend the pellet and add appropriate aliquots of the cell suspension to new culture vessels.
- h) Incubate cultures at 37°C.

**Subcultivation Ratio: A subcultivation ratio of 1:3 - 1:4 is recommended**

**Medium Renewal: Every 2 to 3 days**

## Notes

- a) Upon initial thawing, a higher number of dead cells is observed, which is a normal phenomenon. Significant improvement is seen after adaptation. Once the cells reach a stable state, the number of dead cells decreases after subculturing and the cell growth rate becomes stable.
- b) Ensure that the cell density does not exceed 80%, as overcrowding may lead to reduced viability due to compression.

## Related Products

GCGR	
<a href="#">H_GCGR Reporter CHO-K1 Cell Line</a>	<a href="#">H_GCGR Reporter HEK-293 Cell Line</a>
<a href="#">H_GCGR Reporter HEK-293 DDX35TM Cell Line</a>	<a href="#">Cynomolgus_GCGR HEK-293 Cell Line</a>
<a href="#">H_GCGR CHO-K1 Cell Line</a>	<a href="#">H_GCGR HEK-293 Cell Line</a>
<a href="#">Mouse_GCGR HEK-293 Cell Line</a>	
<a href="#">Anti-H_GCGR hIgG2 Antibody(volagidemab)</a>	
GLP1R	
<a href="#">H_GLP1R Reporter CHO-K1 Cell Line</a>	<a href="#">H_GLP1R Reporter HEK-293 Cell Line</a>
<a href="#">H_GLP1R Reporter HEK-293 DDX35TM Cell Line</a>	<a href="#">H_GLP1R β-Arrestin Reporter CHO-K1 Cell Line</a>
<a href="#">Cynomolgus_GLP1R GIPR CHO-K1 Cell Line</a>	<a href="#">Cynomolgus_GLP1R HEK-293 Cell Line</a>
<a href="#">H_GLP1R CHO-K1 Cell Line</a>	<a href="#">H_GLP1R GIPR CHO-K1 Cell Line</a>
<a href="#">H_GLP1R HEK-293 Cell Line</a>	<a href="#">Mouse_GLP1R GIPR CHO-K1 Cell Line</a>
<a href="#">Mouse_GLP1R HEK-293 Cell Line</a>	<a href="#">Rat_GLP1R HEK-293 Cell Line</a>
<a href="#">Anti-GLP1R hIgG1 Antibody(mAb-36986)</a>	<a href="#">Anti-H_GLP1R hIgG1 Antibody(glutazumab)</a>
FGFR1	
<a href="#">Human FGF-21 Protein; His Tag</a>	
CALCA(CGRP):CALCRL RAMP	
<a href="#">H_CALCRL RAMP1 Reporter HEK-293 Cell Line</a>	<a href="#">H_CALCRL RAMP1 Reporter HEK-293 DDX35TM Cell Line</a>
<a href="#">Cynomolgus_CALCRL RAMP1 HEK-293 Cell Line</a>	<a href="#">H_CALCRL RAMP1 CHO-K1 Cell Line</a>
<a href="#">H_CALCRL RAMP1 HEK-293 Cell Line</a>	
<a href="#">Anti-CALCRL RAMP1 hIgG2 Antibody(Erenumab)</a>	
GIPR	
<a href="#">H_GIPR Reporter CHO-K1 Cell Line</a>	<a href="#">H_GIPR Reporter HEK-293 Cell Line</a>
<a href="#">H_GIPR Reporter HEK-293 DDX35TM Cell Line</a>	<a href="#">Cynomolgus_GIPR CHO-K1 Cell Line</a>
<a href="#">Cynomolgus_GIPR HEK-293 Cell Line</a>	<a href="#">H_GIPR CHO-K1 Cell Line</a>
<a href="#">H_GIPR HEK-293 Cell Line</a>	<a href="#">Mouse_GIPR CHO-K1 Cell Line</a>
<a href="#">Mouse_GIPR HEK-293 Cell Line</a>	
<a href="#">Anti-H_GIPR hIgG1 Antibody(AMG-133)</a>	
ACVR2A:ACTRIIB:Active A	
<a href="#">ACVR2A KO HEK-293 Cell Line</a>	<a href="#">ACVR2B KO HEK-293 Cell Line</a>
<a href="#">Activin A Reporter Cell Line</a>	<a href="#">BRE Reporter 293 Cell Line</a>
<a href="#">H_ACVR2A Reporter Cell Line</a>	<a href="#">H_ACVR2B Reporter Cell Line</a>
<a href="#">H_ACVR2B Reporter DDX35TM Cell Line</a>	<a href="#">H_ACVR2A HEK-293(ACVR2B KO) Cell Line</a>
<a href="#">H_ACVR2B CHO-K1 Cell Line</a>	<a href="#">H_ACVR2B HEK-293(ACVR2A KO) Cell Line</a>
<a href="#">Anti-ACVR2B hIgG1 Antibody(Bimagrumab)</a>	<a href="#">Anti-ACVR2B hIgG1 Antibody(Fab-17G05)</a>
<a href="#">Anti-ACVR2B mIgG2a Antibody(Bimagrumab)</a>	<a href="#">Anti-H_ACVR2B hIgG1 Reference Antibody(Bimbo)</a>
<a href="#">Biotinylated Human ACVR2A Protein; His-Avi Tag</a>	<a href="#">Biotinylated Human ACVR2B Protein; His-Avi Tag</a>
<a href="#">Biotinylated Mouse ACVR2A Protein; His-Avi Tag</a>	<a href="#">Biotinylated Mouse ACVR2B Protein; His-Avi Tag</a>
<a href="#">Human Activin A Protein; His Tag</a>	<a href="#">Human Activin A Protein; His Tag (CHO)</a>

Human Activin B Protein; His Tag	Human ACVR2A Protein; hFc Tag
Human ACVR2A Protein; hFc Tag (Sotatercept)	Human ACVR2A Protein; His Tag
Human ACVR2B Protein; hFc Tag	Human ACVR2B Protein; His Tag
Human latent GDF-8 Protein; His Tag	Mouse ACVR2A Protein; His Tag
Mouse ACVR2B Protein; His Tag	
AMY:CALCR RAMP	
H_CALCR RAMP3(AMY3) Reporter CHO-K1 Cell Line	H_CALCR RAMP3(AMY3) $\beta$ -Arrestin Reporter CHO-K1 Cell Line
H_CALCR Reporter CHO-K1 Cell Line	H_CALCR $\beta$ -Arrestin Reporter CHO-K1 Cell Line
Rat_CALCR RAMP3(AMY3) Reporter COS-7 Cell Line	Rat_CALCR Reporter COS-7 Cell Line
THRB	
H_THR $\beta$ Reporter HEK-293 Cell Line	
MC4R	
H_MC4R Reporter HEK-293 Cell Line	
ASGR1	
H_ASGR1 CHO-K1 Cell Line	H_ASGR1 HEK-293 Cell Line
Cynomolgus ASGR1 Protein; His Tag	Human ASGR1 Protein; His Tag

## License Agreement:

**By purchasing and using this cell line product, the user voluntarily agrees to accept and abide by the following policies:**

- This cell line product is restricted to research use only and shall not be used for any commercial purposes.
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