Genomeditech (Shanghai) Co.,Ltd.
Order: +86 021-68455258/50432826/50432825
Toll-free: +86 400 627 9288
Email: service@genomeditech.com

Product Sheet

H_CASR Reporter CHO-K1 Cell Line

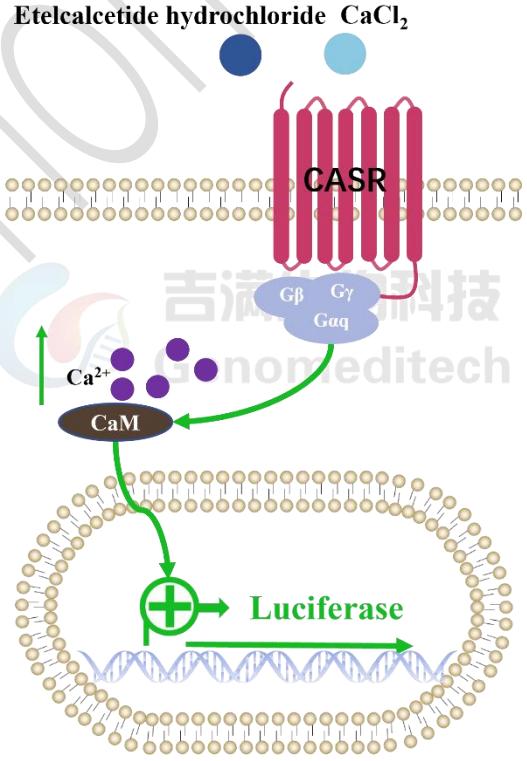
Catalog number: GM-C47275

Version 3.3.1.260717

Calcium-Sensing Receptor (CaSR) is a G protein-coupled receptor encoded by the CASR gene, predominantly expressed in parathyroid chief cells, as well as in renal and bone tissues. It is pivotal for systemic calcium homeostasis, functioning as the primary sensor for extracellular Ca^{2+} to regulate parathyroid hormone (PTH) secretion and bone metabolic balance. CaSR is implicated in various metabolic disorders, including primary hyperparathyroidism and osteoporosis, establishing it as a promising therapeutic target.

The CaSR signaling pathway is initiated upon binding of extracellular Ca^{2+} to the receptor, inducing conformational changes and coupling to heterotrimeric G proteins. This primarily activates the Gq/11 pathway, stimulating phospholipase Cβ (PLCβ) to hydrolyze PIP_2 into IP_3 and DAG, triggering intracellular Ca^{2+} release.

H_CASR Reporter CHO-K1 Cell Line is a clonal stable cell line engineered using lentiviral technology, featuring constitutive expression of the human CASR gene alongside a signal-dependent luciferase reporter gene. Upon binding of agonists to CaSR, the receptor couples to the endogenous Gq proteins, activating downstream effectors that drive luciferase expression. Antagonist antibodies or modulators can inhibit this signal transduction. Quantitative measurement of luciferase activity reflects the activation status of the Gq–Ca^{2+} pathway, providing a robust platform for the in vitro evaluation of CaSR-targeted therapeutics.



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Specifications

Quantity	5E6 Cells per vial,1 mL
Product Format	1 vial of frozen cells
Shipping	Shipped on dry ice
Storage Conditions	Liquid nitrogen immediately upon receipt
Recovery Medium	F12K+10% FBS+1% P.S
Growth medium	F12K+10% FBS+1% P.S+4 μg/mL Blasticidin+4 μg/mL Puromycin
Note	None
Freezing Medium	90% FBS+10% DMSO
Growth properties	Suspension
Growth Conditions	37°C, 5% CO_2
Mycoplasma Testing	The cell line has been screened to confirm the absence of Mycoplasma species.
Safety considerations	Biosafety Level 2
Note	It is recommended to expand the cell culture and store a minimum of 10 vials at an early passage for potential future use.

Materials

Reagent	Manufacturer/Catalogue No.
F12K	BOSTER/PYG0036
Fetal Bovine Serum	ExCell/FSP500
Pen/Strep	Thermo/15140-122
Blasticidin	Genomeditech/GM-040404
Puromycin	Genomeditech/GM-040401
Calcium chloride anhydrous, for cell culture	MCE/HY-Y0406E
Etelcalcetide (hydrochloride)	MCE/HY-P1955A
GMOne-Step 2.0 Luciferase Reporter Gene Assay Kit	Genomeditech/GM-040513

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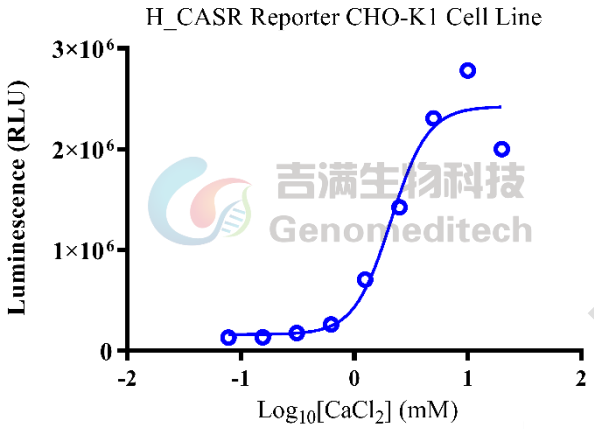
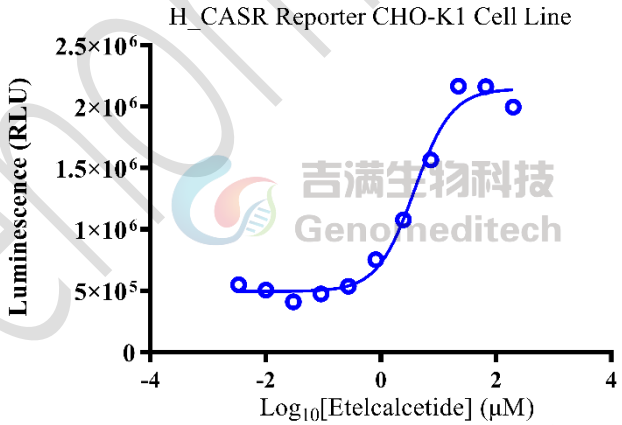
Figures



	H_CASR Reporter CHO-K1
EC50	2.112

Figure 1 | Response to Calcium chloride anhydrous. H_CASR Reporter CHO-K1 Cell Line (Cat. GM-C47275) was seeded at a density of 1E4 cells per well in a 96-well plate and incubated overnight. The next day, serial dilutions of the Calcium chloride anhydrous (MCE/HY-Y0406E) were added to the pre-seeded cells. The mixture was incubated for an additional 16 hours. The firefly luciferase activity was measured using the Luciferase Reporter Assay Kit (Genomeditech). The maximum induction fold was approximately [21.7]. Data are shown by drug molar concentration.



	H_CASR Reporter CHO-K1
EC50	3.849

Figure 2 | Response to Etelcalcetide. H_CASR Reporter CHO-K1 Cell Line (Cat. GM-C47275) was seeded at a density of 1E4 cells per well in a 96-well plate and incubated overnight. The next day, serial dilutions of the Etelcalcetide (hydrochloride) (MCE/HY-P1955A) were added to the pre-seeded cells. The mixture was incubated for an additional 16 hours. The firefly luciferase activity was measured using the Luciferase Reporter Assay Kit (Genomeditech). The maximum induction fold was approximately [4.3]. Data are shown by drug molar concentration.

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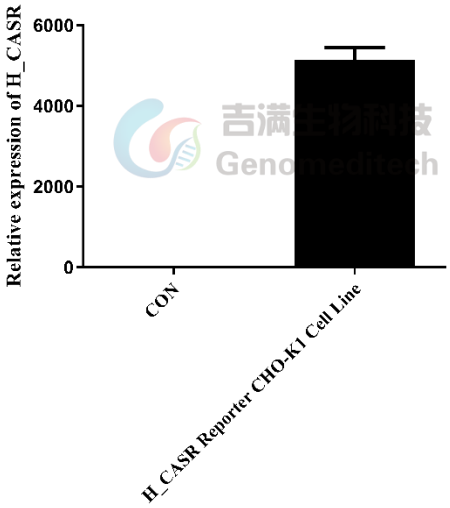

Figure 3 | The mRNA expression levels of H_CASR in the H_CASR Reporter CHO-K1 Cell Line (Cat. GM-C47275) were determined by RT-Qpcr.

Cell Recovery

Recovery Medium: F12K+10% FBS+1% P.S

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

a) Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 - 3 minutes).

b) Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.

c) Transfer the vial contents to a centrifuge tube containing 5.0 mL complete culture medium and spin at approximately 176 x g for 5 minutes. Discard supernatant.

d) Resuspend cell pellet with the recommended recovery medium. And dispense into appropriate culture dishes.

e) Incubate the culture at 37°C in a suitable incubator. A 5% CO_2 in air atmosphere is recommended if using the medium described on this product sheet.

Cell Freezing

Freezing Medium: 90% FBS+10% DMSO

a) Centrifuge at 176 x g for 3 minutes to collect cells.

b) Resuspend the cells in pre-cooled freezing medium and adjust the cell density to 5E6 cells/mL.

c) Aliquot 1 mL into each vial.

d) Place the vial in a controlled-rate freezing container and store at -80°C for at least 1 day, then transfer to liquid nitrogen as soon as possible.

Cell passage

Growth medium: F12K+10% FBS+1% P.S+4 μg/mL Blasticidin+4 μg/mL Puromycin

For the first 1 to 2 passages post-resuscitation, use the recovery medium. Once the cells have stabilized, switch to a growth medium.

a) Remove and discard culture medium.

b) Briefly rinse the cell layer with PBS to remove all traces of serum that contains trypsin inhibitor.

c) Add 1.0 mL of 0.25% (w/v) Trypsin-EDTA solution to dish and observe cells under an inverted microscope until cell layer is dispersed (usually within 2 to 3 minutes at 37°C).

d) Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.

e) Add 2.0 mL of growth medium to mix well and aspirate cells by gently pipetting.

f) After centrifugation, resuspend the pellet and add appropriate aliquots of the cell suspension to new culture vessels.

g) Incubate cultures at 37°C.

Subcultivation Ratio: A subcultivation ratio of 1:4 - 1:5 is recommended

Medium Renewal: Every 2 to 3 days

Notes

a) After the stabilization of the cell condition, there will be fewer dead cells post-passage, the cell growth rate will tend to stabilize, cell morphology will become uniform, and the cells will appear robust.

License Agreement:

By purchasing and using this cell line product, the user voluntarily agrees to accept and abide by the following policies:

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