

# Product Sheet

## H\_HER2 MCF-7 Cell Line

Catalog number: GM-C26434

Version 3.3.1.260126

|                              |   |
|------------------------------|---|
| <b>Description</b>           | H_HER2 MCF-7 Cell Line is a clonal stable MCF-7 cell line that constitutively expresses the human HER2 gene, constructed using lentiviral technology.               |
| <b>Quantity</b>              | 5E6 Cells per vial, 1 mL  |
| <b>Product Format</b>        | 1 vial of frozen cells  |
| <b>Shipping</b>              | Shipped on dry ice  |
| <b>Storage Conditions</b>    | Liquid nitrogen immediately upon receipt  |
| <b>Target</b>                | Human_HER2 & C-3Flag  |
| <b>Gene ID/Uniprot ID</b>    | P04626-1  |
| <b>Host Cell</b>             | MCF-7   |
| <b>Recovery Medium</b>       | MEM(Gibco)+20% FBS+1% P.S+0.01 mg/mL Bovine Insulin   |
| <b>Growth medium</b>         | MEM(Gibco)+10% FBS+1% P.S+0.01 mg/mL Bovine Insulin+0.75 µg/mL Puromycin  |
| <b>Note</b>                  | Cells should be cultured using Gibco/11095080 MEM medium or Growth medium from Genomeditech. The serum should be Cegrogen biotech/A0500-3010 or sourced from Gibco. |
| <b>Freezing Medium</b>       | 90% FBS+10% DMSO  |
| <b>Growth properties</b>     | Adherent  |
| <b>Growth Conditions</b>     | 37°C, 5% CO <sub>2</sub>  |
| <b>Mycoplasma Testing</b>    | The cell line has been screened to confirm the absence of Mycoplasma species.   |
| <b>Safety considerations</b> | Biosafety Level 2   |
| <b>Note</b>                  | It is recommended to expand the cell culture and store a minimum of 10 vials at an early passage for potential future use.  |

## Materials

| Reagent                                  | Manufacturer/Catalogue No.               |
|--|--|
| MEM                                      | gibco/11095-080                          |
| Fetal Bovine Serum                       | ExCell/FSP500                            |
| Insulin(From Bovine)                     | YEASEN/40107ES25                         |
| Puromycin                                | Genomeditech/ <a href="#">GM-040401</a>  |
| Anti-H_HER2 hIgG1 Antibody(Margetuximab) | Genomeditech/ <a href="#">GM-49468AB</a> |

## Figures

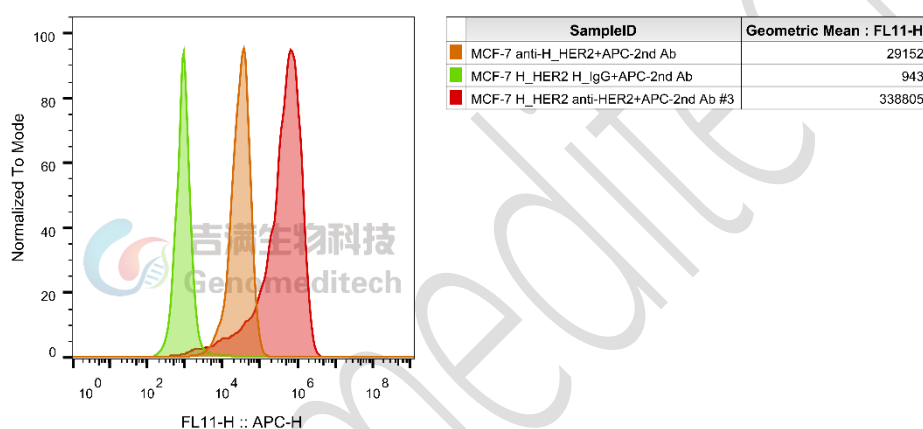


Figure 1 | H\_HER2 MCF-7 Cell Line (Cat. GM-C26434) was determined by flow cytometry using Anti-H\_HER2 hIgG1 Antibody(Margetuximab) (Cat. [GM-49468AB](#)).

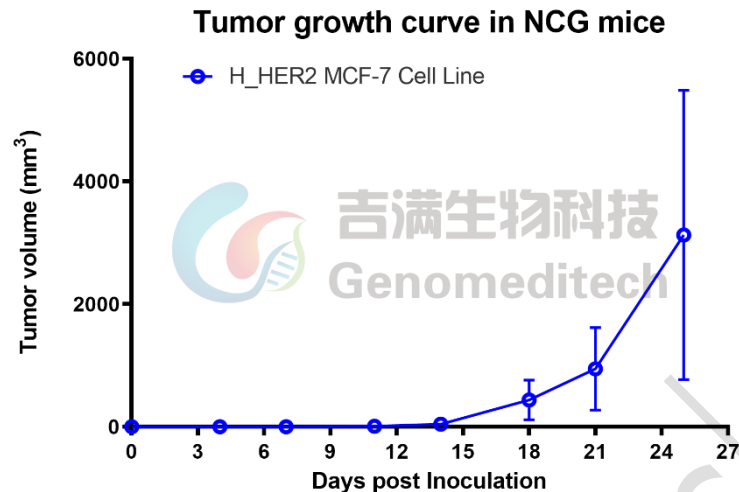


Figure 2 | Tumor growth curves of H\_HER2 MCF-7 in NCG mice. H\_HER2 MCF-7 cells ( $1 \times 10^6$  per mouse) were subcutaneously inoculated into NCG mice (female, 8 weeks old,  $n = 3$ ). Tumor volume was measured twice per week and is presented as mean  $\pm$  SEM.

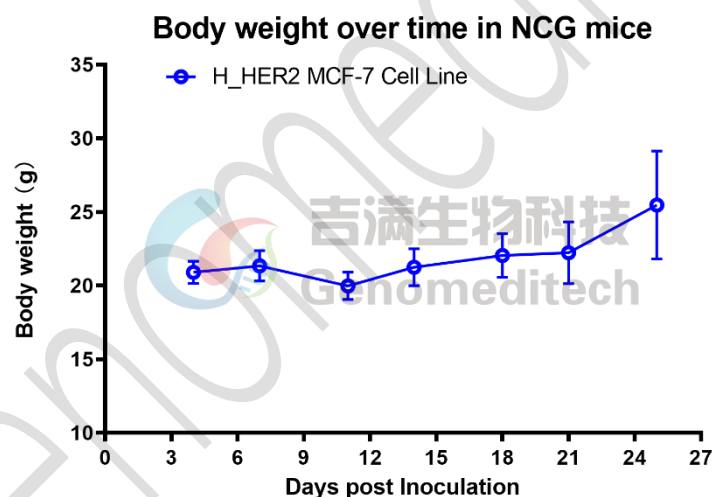


Figure 3 | Body weight changes after implantation of H\_HER2 MCF-7 in NCG mice. Under the same conditions, body weight was measured twice per week and is presented as mean  $\pm$  SEM.

## Cell Recovery

Recovery Medium: MEM(Gibco)+20% FBS+1% P.S+0.01 mg/mL Bovine Insulin

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at  $-70^{\circ}\text{C}$ . Storage at  $-70^{\circ}\text{C}$  will result in loss of viability.

- Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 - 3 minutes).
- Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
- Transfer the vial contents to a centrifuge tube containing 5.0 mL complete culture medium and spin at approximately 176 x g for 5 minutes. Discard supernatant.
- Resuspend cell pellet with the recommended recovery medium. And dispense into appropriate culture dishes.
- Incubate the culture at 37°C in a suitable incubator. A 5% CO<sub>2</sub> in air atmosphere is recommended if using the medium described on this product sheet.

## Cell Freezing

Freezing Medium: 90% FBS+10% DMSO

- Centrifuge at 176 x g for 3 minutes to collect cells.
- Resuspend the cells in pre-cooled freezing medium and adjust the cell density to 5E6 cells/mL.
- Aliquot 1 mL into each vial.
- Place the vial in a controlled-rate freezing container and store at -80°C for at least 1 day, then transfer to liquid nitrogen as soon as possible.

## Cell passage

Growth medium: MEM(Gibco)+10% FBS+1% P.S+0.01 mg/mL Bovine Insulin+0.75 µg/mL Puromycin

For the first 1 to 3 passages post-resuscitation, use the recovery medium. Once the cells have stabilized, switch to a growth medium.

- Remove and discard culture medium.
- Briefly rinse the cell layer with PBS to remove all traces of serum that contains trypsin inhibitor.
- Add 1.0 mL of 0.25% (w/v) Trypsin-EDTA solution to dish and observe cells under an inverted microscope until cell layer is dispersed (usually within 1 to 2 minutes at 37°C).
- Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.
- Add 2.0 mL of growth medium to mix well and aspirate cells by gently pipetting.
- After centrifugation, resuspend the pellet and add appropriate aliquots of the cell suspension to new culture vessels.
- Incubate cultures at 37°C.

**Subcultivation Ratio: A subcultivation ratio of 1:2 - 1:3 is recommended**

**Medium Renewal: Every 3 to 5 days**

## Notes

- After cell resuscitation, the first few generations exhibit slow adherence, which is a normal phenomenon. Adherent cells can be observed 2 - 3 days after resuscitation. Cells exhibit density dependence, growing very slowly at low densities. Once their condition has stabilized, their growth rate tends to become consistent.

- b) Due to the potentially long subcultivation intervals after the initial resuscitation, it is necessary to add fresh medium or replace it entirely if no handling occurs for 2 - 3 days.
- c) During cell culture, some floating cells may be present. Viable suspended cells can be re-cultured in the flask by centrifugation and re-seeding (specifically observe during passaging; if the proportion is small, they can be discarded directly).
- d) Cells should be cultured using Gibco/11095080 MEM medium or complete media from Geomeditech. Serum should be the same as specified in the instructions or use Gibco serum.

## Sequence

HER2(ERBB2)-3Flag [P04626-1](#)

MELAALCRWGLLLALLPPGAASTQVCTGTDMKLRASPETHLDMLRHL YQGCQVVQGNLELTYLPTNASL  
SFLQDIQEVQGYVLIAHNQVRQVPLQRLRIVRGTLFEDNYALAVLDNGDPLNNTTPVTGASPGGLRELQLR  
SLTEILKGGVLIQRNPQLCYQDTILWKDIFHKNNQLALTLIDTNRSRACHPCSPMCKGSRWGSESSEDCQSLT  
RTVCAGGCARCKGPLPTDCCHEQCAAGCTGPKHSDCLACLFHNHSGICELHCPALVTYNTDTFESMPNPEGR  
YTFGASCVTACPYNYLSTDVGSCTLVCPHNQEVTAEDGTQRCEKCSKPCARVCYGLGMEHLREVRAVTS  
NIQEFAGCKKIFGSLAFLPESFDGDPASNTAPLQPEQLQVFETLEEITGYLYISAWPDSLPLSVFQNLQVIRGRI  
LHNGAYSLTLQGLGISWLGLRSLRELGSGLALIHNTHLCFVHTVPWDQLFRNPHQALLHTANRPEDECVGE  
GLACHQLCARGHCWGPGPTQCVNCSQFLRGQECVEECRVLQGLPREYVNARHCLPCHPECQPQNGSVTCFG  
PEADQCVACAHYKDPFFCVARCPSGVKPDL SYMPIWKFPDEEGACQPCINCTHSCVDLDDKGCPAEQRAS  
LTSIISAVVGILLVVVLGVVFGILIKRRQKIRKYTMRRLLQETELVEPLTPSGAMPNQAQMRILKETELRKVK  
VLGSGAFGTVYKGIWIPDGENVKIPVAIKVLRNTSPKANKEILDEAYVMAGVGSPYVSRLLGICLTSTVQLV  
TQLMPYGCLLDHVRENRLGSLQDLLNWCMIKAGMSYLEDVRLVHRDLAARNVLVKSPNHVKITDFGLA  
RLLDIDETEHADGGKVPKWMALLESILRRRFTHQSDVWSYGVTVWELMTFGAKPYDGIPAREIPDLLEKGE  
RLPQPPICTIDVYMIMVKCWMIDSECRPRFREL VSEFSRMARDPQRFVVIQNE DLGPASPLDSTFYRSLLEDD  
MGDLVDAEEYLVPQQGFFCPDPAPGAGGMVHHRHRSSTRSGGDLTLGLEPSEEEAPRSPLAPSEGAGSDV  
FDGDLGMGAAGKLQSLPTHDPSPQLRYSDEPTVPLPSETDGYVAPLTCSPQPEYVNQPDVRPQPPSPREGPLP  
AARPAGATLERPKTLSPGKNGVVKDVFAFGGAVENPEYLPQGGGAAPQPHPPAFSPAFDNLYYWDQDPPE  
RGAPPSTFKGTPTAENPEYLGDLVPVLESRTTRPGPSGMDYKDHDGDYKDHDIDYKDDDDK

## Related Products

| EGFR:HER2:HER3:HER4                                      |   |
|--|---|
| <a href="#">H_EGFR Reporter Cell Line</a>                | <a href="#">H_HER2 HER4 Reporter HEK-293 Cell Line</a>  |
| <a href="#">Cynomolgus_EGFR CHO-K1 Cell Line</a>         | <a href="#">Cynomolgus_ERBB3(HER3) CHO-K1 Cell Line</a> |
| <a href="#">Cynomolgus_ERBB3(HER3) HEK-293 Cell Line</a> | <a href="#">Cynomolgus_HER2(ERBB2) CHO-K1 Cell Line</a> |
| <a href="#">H_EGFR CHO-K1 Cell Line</a>                  | <a href="#">H_EGFR HEK-293 Cell Line</a>                |
| <a href="#">H_EGFRvIII CHO-K1 Cell Line</a>              | <a href="#">H_ERBB3(HER3) CHO-K1 Cell Line</a>          |
| <a href="#">H_ERBB3(HER3) HEK-293 Cell Line</a>          | <a href="#">H_ERBB3(HER3) MC38 Cell Line</a>            |
| <a href="#">H_HER2 EMT6 Cell Line</a>                    | <a href="#">H_HER2 HER3 MC38 Cell Line</a>              |
| <a href="#">H_HER2(ERBB2) CHO-K1 Cell Line</a>           | <a href="#">H_HER2(ERBB2) CT26 Cell Line</a>            |

|   |  |
|---|--|
| H_HER2(ERBB2) LLC1 Cell Line                        | H_HER2(ERBB2) MC38 Cell Line                 |
| Mouse_HER3(ERBB3) CHO-K1 Cell Line                  | H_EGFR LGR5 CT26 Cell Line                   |
| H_EGFR LGR5 HEK-293 Cell Line                       |  |
| Anti-EGFR hIgG1 Reference Antibody (Cetbio)         | Anti-EGFR hIgG1 Reference Antibody (Necbio)  |
| Anti-ERBB3(HER3) hIgG1 Reference Antibody(Patribio) | Anti-H_EGFR hIgG1 Antibody(Necitumumab)      |
| Anti-H_ERBB3(HER3) hIgG1 Antibody(Barecetamab)      | Anti-H_HER2 hIgG1 Antibody(Margetuximab)     |
| Anti-HER2 hIgG1 Reference Antibody(Marbio)          | Anti-HER2 hIgG1 Reference Antibody(Trasbio)  |
| Biotinylated Cynomolgus EGFR Protein; His-Avi Tag   | Biotinylated Human EGFR Protein; His-Avi Tag |
| Biotinylated Human HER3 Protein; His-Avi Tag        | Cynomolgus HER2 Protein; His Tag             |
| Human EGFR Protein; His Tag                         | Human HER2 Protein; His Tag                  |
| Human HER3 Protein; His Tag                         | Mouse HER3 Protein; His Tag                  |

## License Agreement:

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