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Product Sheet

Mouse PDL1 VEGFA KO CT26 Cell Line

Catalog number: GM-C45469

Version 3.3.1.260428

Description	Mouse PDL1 VEGFA KO CT26 Cell Line is a clonal stable cell line derived from CT26 cells with a knockout of mouse PDL1 and VEGFA.
Quantity	5E6 Cells per vial,1 mL
Product Format	1 vial of frozen cells
Shipping	Shipped on dry ice
Storage Conditions	Liquid nitrogen immediately upon receipt

Target	Mouse PDL1 VEGFA KO
Gene ID/Uniprot ID	/
Host Cell	CT26 Cell Line

Recovery Medium	RPMI 1640+10% FBS+1% P.S
Growth medium	RPMI 1640+10% FBS+1% P.S
Note	None
Freezing Medium	90% FBS+10% DMSO
Growth properties	Adherent
Growth Conditions	37°C, 5% CO_2

Mycoplasma Testing	The cell line has been screened to confirm the absence of Mycoplasma species.
Safety considerations	Biosafety Level 1
Note	It is recommended to expand the cell culture and store a minimum of 10 vials at an early passage for potential future use.

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Materials

Reagent	Manufacturer/Catalogue No.
RPMI 1640	gibco/C11875500BT
Fetal Bovine Serum	ExCell/FSP500
Pen/Strep	Thermo/15140-122
Mouse IFNG/Interferon Gamma Protein	Sino Biological/50709-MNAH
PE anti-mouse CD274 (B7-H1, PD-L1) Antibody	BioLegend/124307
Mouse VEGF ELISA Kit	Yeasen/98020ES48

Figures

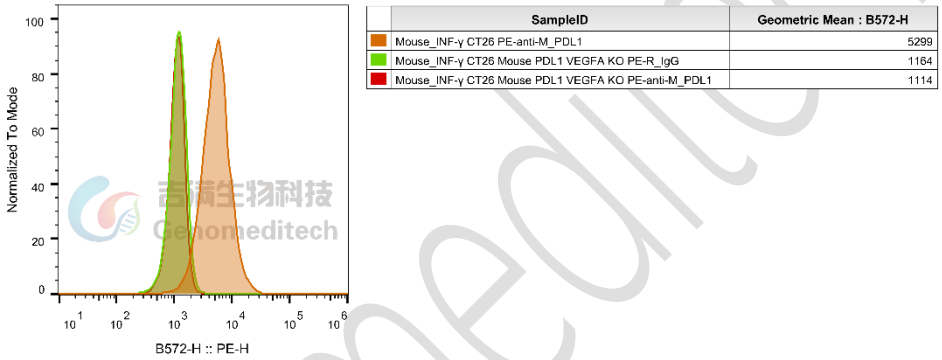

Figure 1 | Mouse PDL1 VEGFA KO CT26 Cell Line (Cat. GM-C45469) was determined by flow cytometry using PE anti-mouse CD274 (B7-H1, PD-L1) Antibody (BioLegend/124307).

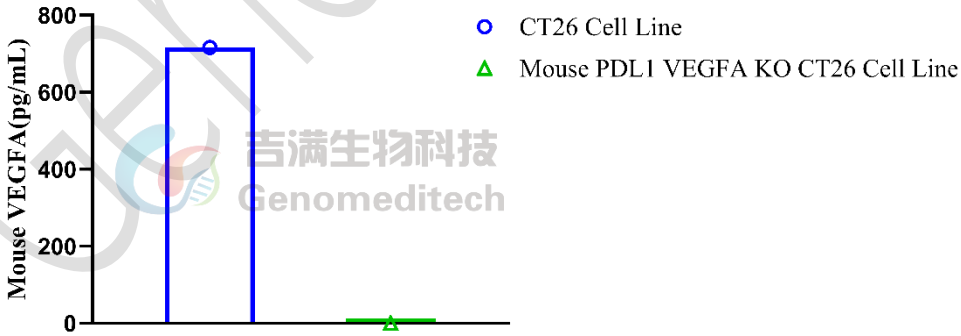

Figure 2 | Mouse PDL1 VEGFA KO CT26 Cell Line (Cat. GM-C45469) was determined by Elisa using Mouse VEGF ELISA Kit (Yeasen/98020ES48).

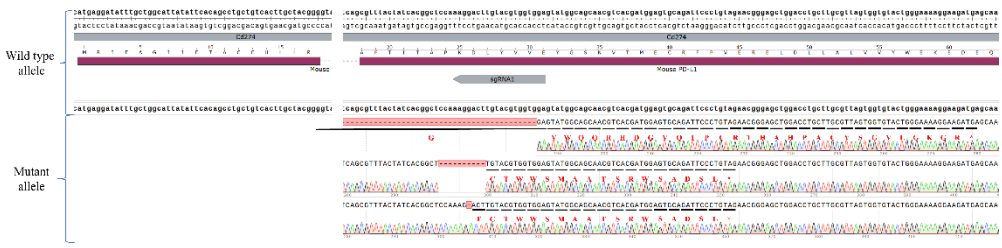

Figure 3 | The Sanger sequencing of the Mouse PDL1 VEGFA KO CT26 Cell Line showed successful knockout of PDL1.

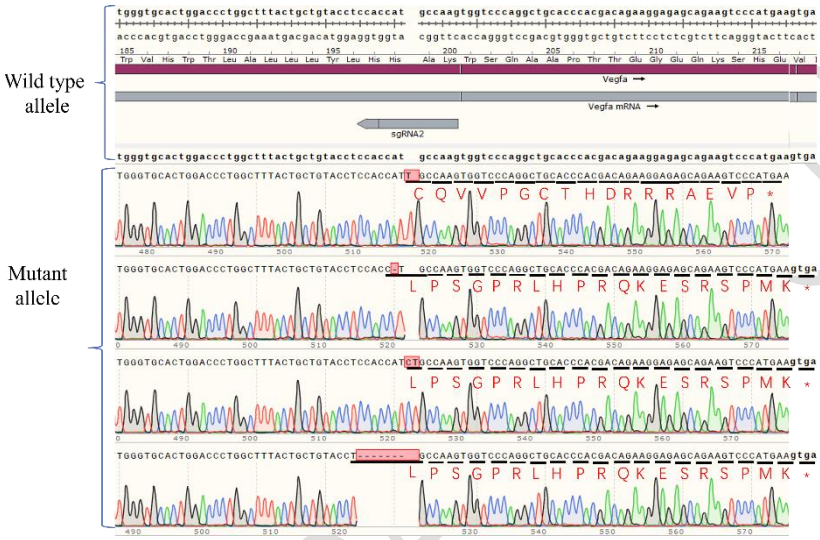

Figure 4 | The Sanger sequencing of the Mouse PDL1 VEGFA KO CT26 Cell Line showed successful knockout of VEGFA.

Cell Recovery

Recovery Medium: RPMI 1640+10% FBS+1% P.S

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

a) Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 - 3 minutes).

b) Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.

c) Transfer the vial contents to a centrifuge tube containing 5.0 mL complete culture medium and spin at approximately 176 x g for 5 minutes. Discard supernatant.

d) Resuspend cell pellet with the recommended recovery medium. And dispense into appropriate culture dishes.

e) Incubate the culture at 37°C in a suitable incubator. A 5% CO_2 in air atmosphere is recommended if using the medium described on this product sheet.

Cell Freezing

Freezing Medium: 90% FBS+10% DMSO

a) Centrifuge at 176 x g for 3 minutes to collect cells.

b) Resuspend the cells in pre-cooled freezing medium and adjust the cell density to 5E6 cells/mL.

c) Aliquot 1 mL into each vial.

d) Place the vial in a controlled-rate freezing container and store at -80°C for at least 1 day, then transfer to liquid nitrogen as soon as possible.

Cell passage

Growth medium: RPMI 1640+10% FBS+1% P.S

For the first 1 to 2 passages post-resuscitation, use the recovery medium. Once the cells have stabilized, switch to a growth medium.

a) Remove and discard culture medium.

b) Briefly rinse the cell layer with PBS to remove all traces of serum that contains trypsin inhibitor.

c) Add 1.0 mL of 0.25% (w/v) Trypsin-EDTA solution to dish and observe cells under an inverted microscope until cell layer is dispersed (usually within 30 to 60 seconds at 37°C).

d) Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.

e) Add 2.0 mL of growth medium to mix well and aspirate cells by gently pipetting.

f) After centrifugation, resuspend the pellet and add appropriate aliquots of the cell suspension to new culture vessels.

g) Incubate cultures at 37°C.

Subcultivation Ratio: A subcultivation ratio of 1:3 - 1:5 is recommended

Medium Renewal: Every 2 to 3 days

Notes

a) After the stabilization of the cell condition, there will be fewer dead cells post-passage, the cell growth rate will tend to stabilize, cell morphology will become uniform, and the cells will appear robust.

Related Products

VEGF:VEGFR	
H_VEGF Reporter 293 Cell Line	H_VEGF Reporter 293 DDX35TM Cell Line
H_VEGFR1 CHO-K1 Cell Line	H_VEGFR1 HEK-293 Cell Line
Membrane Bound H_VEGF165 CHO-K1 Cell Line	
Anti-mouse VEGFR-2 mIgG2a Antibody(DC101)	Anti-mouse VEGFR-2 RIgG1 Antibody(DC101)
Anti-Mouse_PD1×VEGF hIgG1 Bispecific Antibody	Anti-VEGF hIgG1 Antibody(Bevacizumab)
Anti-VEGF hIgG1 Reference Antibody (Bevbio)	Anti-VEGF×PD1 hIgG1 Reference Antibody (Ivobio)
Anti-VEGF×PD-L1 hIgG1 Bispecific Antibody (Pumibio)	Anti-VEGFR-1 hIgG1 Antibody (Icrucumab)

Anti-VEGFR2 hIgG1 Antibody(ramucirumab)	Anti-VEGF×PD-L1 hIgG1 Bispecific Antibody (pumitamig)
Biotinylated Human VEGF110 Protein; His-Avi Tag	Biotinylated Human VEGF121 Protein; His-Avi Tag
Biotinylated Human VEGFR2 Protein; His-Avi Tag	Human VEGF110 Protein; His Tag
Human VEGF121 Protein; His Tag	Human VEGF165 Protein; His Tag
Human VEGFR1 Protein; His Tag	Human VEGFR2 Protein; hFc Tag
Human VEGFR2 Protein; His Tag	Mouse VEGF120 Protein; His Tag
Mouse VEGF164 Protein; His Tag	Mouse VEGFR2 Protein; His Tag

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