

# Product Sheet

## H\_IL12 Reporter 293 Cell Line

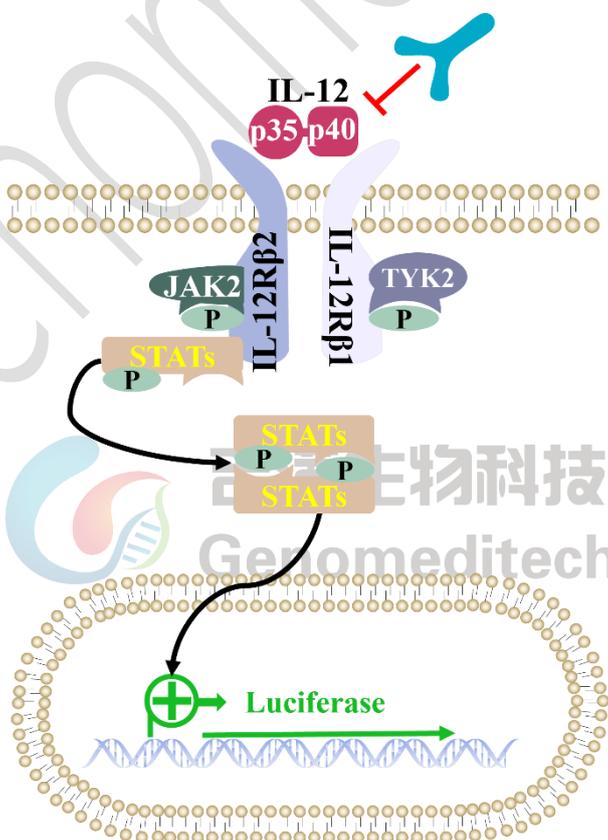
Catalog number: GM-C19224

Version 3.3.1.260201

Interleukin-12 (IL-12) is a cytokine produced by macrophages and dendritic cells, consisting of two subunits: p35 and p40, which form the active dimer IL-12p70. It is essential for immune responses, particularly in activating and proliferating T cells and natural killer (NK) cells, and enhances the production of interferon-gamma (IFN- $\gamma$ ) to strengthen immunity against viruses and tumors.

IL-12 binds to its receptor IL-12R, which has two subunits (IL-12R $\beta$ 1 and IL-12R $\beta$ 2), activating the tyrosine kinases JAK2 and TYK2. This activation leads to the phosphorylation of the STAT4 transcription factor, which then regulates the expression of genes, including IFN- $\gamma$  production. This pathway promotes T cell differentiation into Th1 cells and enhances NK cell cytotoxicity, boosting the immune response against pathogens and tumors.

H\_IL12 Reporter 293 Cell Line is a clonal stable HEK-293 cell line constructed using lentiviral technology, constitutive expression of the IL-12R $\beta$ 1 and IL-12R $\beta$ 2 gene, along with signal-dependent expression of a luciferase reporter gene. When IL12 binds to IL-12R, it activates downstream signaling pathways, leading to the expression of luciferase. Blockade antibodies can inhibit this signal transmission. The luciferase activity measurement indicates the activation level of the signaling pathway and can thus be used to evaluate the in vitro effects of drugs related to IL12.



## Specifications

<b>Quantity</b>	5E6 Cells per vial, 1 mL
<b>Product Format</b>	1 vial of frozen cells
<b>Shipping</b>	Shipped on dry ice
<b>Storage Conditions</b>	Liquid nitrogen immediately upon receipt
<b>Recovery Medium</b>	DMEM+10% FBS+1% P.S
<b>Growth medium</b>	DMEM+10% FBS+1% P.S+4 µg/mL Blasticidin+400 µg/mL G418+125 µg/mL Hygromycin+0.75 µg/mL Puromycin
<b>Note</b>	None
<b>Freezing Medium</b>	90% FBS+10% DMSO
<b>Growth properties</b>	Adherent
<b>Growth Conditions</b>	37°C, 5% CO <sub>2</sub>
<b>Mycoplasma Testing</b>	The cell line has been screened to confirm the absence of Mycoplasma species.
<b>Safety considerations</b>	Biosafety Level 2
<b>Note</b>	It is recommended to expand the cell culture and store a minimum of 10 vials at an early passage for potential future use.

## Materials

Reagent	Manufacturer/Catalogue No.
DMEM	Gibco/C11995500BT
Fetal Bovine Serum	ExCell/FSP500
Pen/Strep	Thermo/15140-122
Blasticidin	Genomeditech/ <a href="#">GM-040404</a>
G418	Genomeditech/ <a href="#">GM-040402</a>
Hygromycin	Genomeditech/ <a href="#">GM-040403</a>
Puromycin	Genomeditech/ <a href="#">GM-040401</a>
Recombinant Human IL-12 Protein (His Tag)	Sino Biological/CT011-H08H
Recombinant Human IL-12	Novoprotein/CI58
Anti-IL-12/23(p40) hIgG1 Reference Antibody (Ustebio)	Genomeditech/ <a href="#">GM-87870MAB</a>
GMOne-Step 2.0 Luciferase Reporter Gene Assay Kit	Genomeditech/ <a href="#">GM-040513</a>

## Figures

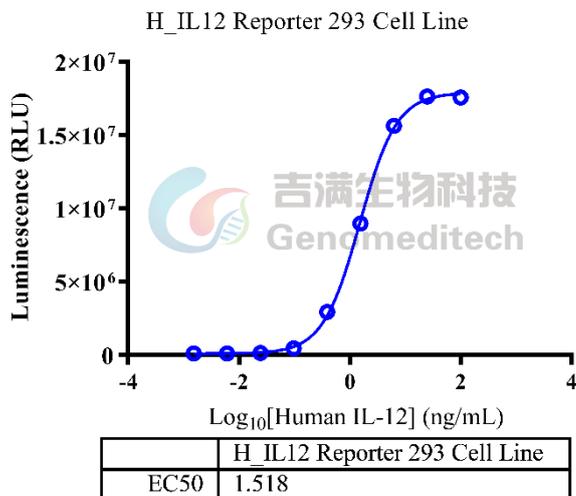


Figure 1 | Response to Recombinant Human IL-12 Protein. The H\_IL12 Reporter 293 Cell Line (Cat. GM-C19224) at a concentration of 1.5E4 cells/well (96-well format) was stimulated with serial dilutions of Recombinant Human IL-12 Protein (Novoprotein/CI58) in assay buffer (DMEM+1% FBS+1% P.S) for 7 hours. The firefly luciferase activity was measured using the Luciferase Reporter Assay Kit (Genomeditech). The maximum induction fold was approximately [138.6]. Data are shown by drug mass concentration.

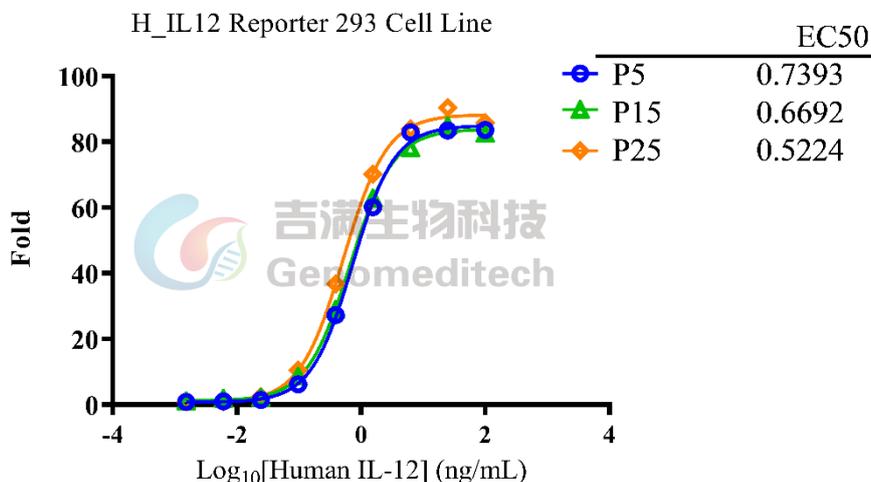


Figure 2 | The passage stability of response to Recombinant Human IL-12 Protein. The passage 5, 15 and 25 of H\_IL12 Reporter 293 Cell Line (Cat. GM-C19224) at a concentration of 1.5E4 cells/well (96-well format) were stimulated with serial dilutions of Recombinant Human IL-12 Protein (Sino Biological/CT011-H08H) in assay buffer (DMEM + 1% FBS + 1% P.S) for 7 hours. The firefly luciferase activity was measured using the Luciferase Reporter Assay Kit (Genomeditech). Data are shown by drug mass concentration.

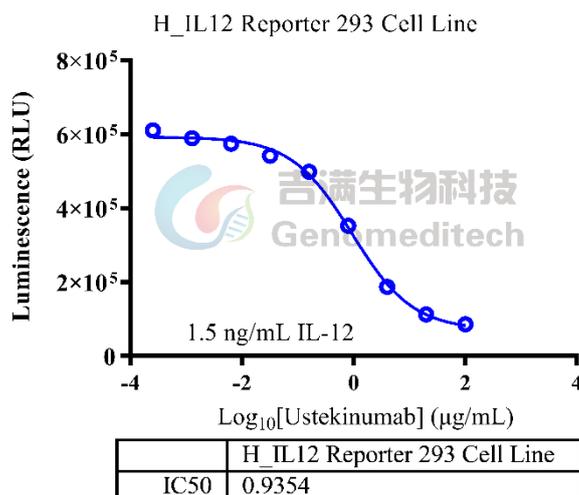


Figure 3 | Response to Anti-IL-12/23(p40) hIgG1 Reference Antibody (Ustebio). Serial dilutions of Anti-IL-12/23(p40) hIgG1 Reference Antibody (Ustebio) (Cat. [GM-87870MAB](#)) was incubated with 0.15 ng/well of Recombinant Human IL-12 (Novoprotein/CI58) for 1 hour in assay buffer (DMEM+1% FBS+1% P.S). After pre-incubation, add the mixture to the H\_IL12 Reporter 293 Cell Line (Cat. GM-C19224) at a density of 1.5E4 cells/well in a 96-well format, and incubate for 6 hours. Firefly luciferase activity was then measured using the Luciferase Reporter Assay Kit (Genomeditech). The results indicated maximum blocking folds of approximately [7.2]. Data are shown by drug mass concentration.

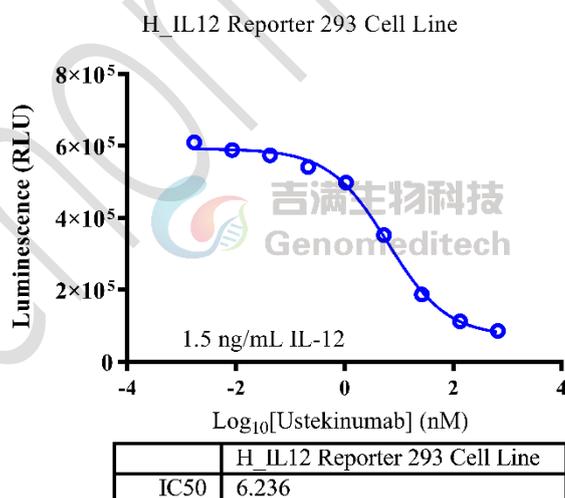


Figure 4 | Response to Anti-IL-12/23(p40) hIgG1 Reference Antibody (Ustebio). Serial dilutions of Anti-IL-12/23(p40) hIgG1 Reference Antibody (Ustebio) (Cat. [GM-87870MAB](#)) was incubated with 0.15 ng/well of Recombinant Human IL-12 (Novoprotein/CI58) for 1 hour in assay buffer (DMEM+1% FBS+1% P.S). After pre-incubation, add the mixture to the H\_IL12 Reporter 293 Cell Line (Cat. GM-C19224) at a density of 1.5E4 cells/well in a 96-well format, and incubate for 6 hours. Firefly luciferase activity was then measured using the Luciferase

Reporter Assay Kit (Genomeditech). The results indicated maximum blocking folds of approximately [7.2]. Data are shown by drug molar concentration.

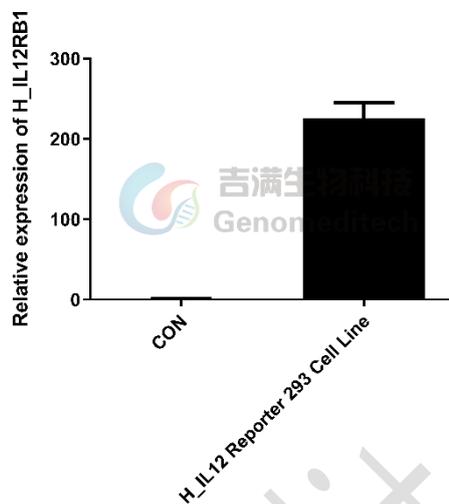


Figure 5 | The mRNA expression levels of IL12RB1 in the H\_IL12 Reporter 293 Cell Line (Cat. GM-C19224) were determined by RT-qPCR.

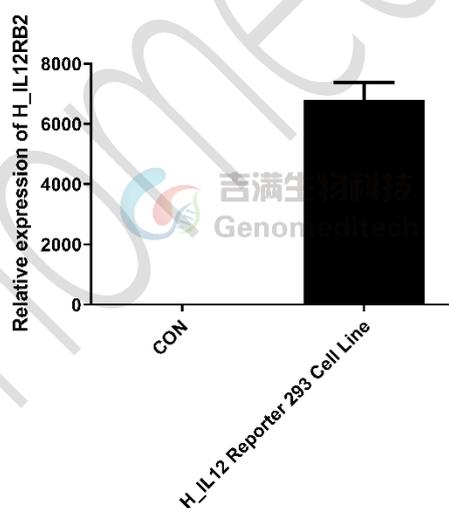


Figure 6 | The mRNA expression levels of IL12RB2 in the H\_IL12 Reporter 293 Cell Line (Cat. GM-C19224) were determined by RT-qPCR.

## Cell Recovery

Recovery Medium: DMEM+10% FBS+1% P.S

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at  $-70^{\circ}\text{C}$ . Storage at  $-70^{\circ}\text{C}$  will result in loss of viability.

- a) Thaw the vial by gentle agitation in a  $37^{\circ}\text{C}$  water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 - 3 minutes).
- b) Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
- c) Transfer the vial contents to a centrifuge tube containing 5.0 mL complete culture medium and spin at approximately  $176 \times g$  for 5 minutes. Discard supernatant.
- d) Resuspend cell pellet with the recommended recovery medium. And dispense into appropriate culture dishes.
- e) Incubate the culture at  $37^{\circ}\text{C}$  in a suitable incubator. A 5%  $\text{CO}_2$  in air atmosphere is recommended if using the medium described on this product sheet.

## Cell Freezing

Freezing Medium: 90% FBS+10% DMSO

- a) Centrifuge at  $176 \times g$  for 3 minutes to collect cells.
- b) Resuspend the cells in pre-cooled freezing medium and adjust the cell density to  $5 \times 10^6$  cells/mL.
- c) Aliquot 1 mL into each vial.
- d) Place the vial in a controlled-rate freezing container and store at  $-80^{\circ}\text{C}$  for at least 1 day, then transfer to liquid nitrogen as soon as possible.

## Cell passage

Growth medium: DMEM+10% FBS+1% P.S+4  $\mu\text{g/mL}$  Blasticidin+400  $\mu\text{g/mL}$  G418+125  $\mu\text{g/mL}$  Hygromycin+0.75  $\mu\text{g/mL}$  Puromycin

For the first 1 to 2 passages post-resuscitation, use the recovery medium. Once the cells have stabilized, switch to a growth medium.

- a) Subculturing is necessary when the cell density reaches 80%. It is recommended to perform subculturing at a ratio of 1:3 to 1:4 every 2-3 days. Ensure that the density does not exceed 80%, as overcrowding can lead to reduced viability due to compression.
- b) Remove and discard culture medium.
- c) Briefly rinse the cell layer with PBS to remove all traces of serum that contains trypsin inhibitor.
- d) Add 1.0 mL of 0.25% (w/v) Trypsin-EDTA solution to dish and observe cells under an inverted microscope until cell layer is dispersed (usually within 30 to 60 seconds at  $37^{\circ}\text{C}$ ).
- e) Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at  $37^{\circ}\text{C}$  to facilitate dispersal.
- f) Add 2.0 mL of growth medium to mix well and aspirate cells by gently pipetting.
- g) After centrifugation, resuspend the pellet and add appropriate aliquots of the cell suspension to new culture vessels.

h) Incubate cultures at 37°C.

**Subcultivation Ratio: A subcultivation ratio of 1:3 - 1:4 is recommended**

**Medium Renewal: Every 2 to 3 days**

## Notes

- Upon initial thawing, a higher number of dead cells is observed, which is a normal phenomenon. Significant improvement is seen after adaptation. Once the cells reach a stable state, the number of dead cells decreases after subculturing and the cell growth rate becomes stable.
- Ensure that the cell density does not exceed 80%, as overcrowding may lead to reduced viability due to compression.

## Related Products

IL-12	
<a href="#">H_IL12 Reporter 293 DDX35TM Cell Line</a>	<a href="#">H_IL12RB1 HEK-293 Cell Line</a>
<a href="#">Anti-IL-12/23(p40) hIgG1 antibody(Ustekinumab)</a>	<a href="#">Anti-IL-12/23(p40) hIgG1 Reference Antibody (Ustebio)</a>
<a href="#">Biotinylated Human IL-23A&amp;IL-12B Heterodimer Protein; His-Avi Tag</a>	<a href="#">Cynomolgus IL-23A &amp; Human IL-12B Heterodimer Protein; His Tag</a>
<a href="#">Cynomolgus IL-23A &amp; Mouse IL-12B Heterodimer Protein; His Tag</a>	<a href="#">Human IL-12B Protein; His Tag</a>
<a href="#">Human IL-23A &amp; Mouse IL-12B Heterodimer Protein; His Tag</a>	<a href="#">Human IL-23A&amp;IL-12B Heterodimer Protein; His Tag</a>
<a href="#">Mouse IL-23A&amp;IL-12B Heterodimer Protein; His Tag</a>	

## License Agreement:

**By purchasing and using this cell line product, the user voluntarily agrees to accept and abide by the following policies:**

- This cell line product is restricted to research use only and shall not be used for any commercial purposes.
- This product is strictly prohibited from being used in the diagnosis or treatment of human or animal diseases, and shall not be directly used in experiments involving humans.
- Users and their contractors engaged for their benefit may use this material and its derivatives only within the agreed research scope; modification of the material is not permitted, nor may it be distributed, sold, transferred, or otherwise provided to any other entity (including affiliates).
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